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(54) Title: ACTIVIN RECEPTORS-LIKE KINASE (ALK), BELONGING TO THE TGF RECEPTOR FAMILY AND/OR TO THE BMP RECEPTOR FAMILY

(57) Abstract

Novel serine/threonine receptor proteins and BMP receptor proteins are disclosed, as well as DNA molecules encoding said proteins and methods of using the receptor proteins. Further disclosed are truncated BMP receptor proteins and molecules which act as ligands to said BMP receptor proteins.

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ACTIVIN RECEPTORS-LIKE KINASE (ALK), BELONGING TO THE TGF RECEPTOR FAMILY AND/OR TO THE BMP RECEPTOR FAMILY

FIELD OF THE INVENTION

The present invention relates to novel serine/threonine kinase receptor proteins, including a novel family of receptor proteins to bone morphogenetic proteins (BMPs). More particularly, the present invention relates to receptor proteins which are able to bind to BMPs, including BMP-2 and BMP-4. The present invention further relates to methods of isolating novel BMP receptor proteins using newly identified DNA fragments as probes for isolating such proteins.

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BACKGROUND OF THE INVENTION

Bone morphogenetic proteins (BMPs) are a family of proteins which have been identified as having the ability to induce the formation of bone and cartilage in tissue extracts. BMPs are a subfamily within the $TGF-\beta$ superfamily. BMPs have multiple therapeutic uses, including a wide variety of settings where bone has been lost through physicological or traumatic processes.

The TGF- β superfamily of proteins have been shown to bind to serine/threonine kinase receptors. Massague, Cell, 69:1067-1070 (1992); Attisano et al., Cell 68:97-108 (1992); Lin et al., Cell, 68:775-785 (1992); Wang et al., Cell 67:797-805 (1991). Similarly, activin receptors have been isolated and characterized as a predicted transmembrane serine kinase. Mathews et al., Cell 65:973-982 (1991); Nakamura et al., J. Biol. Chem. 267:18924-18928 (1992). Ebner et al., Science, 260:1344-1348 (1993) describe the existence of Type I and Type II TGF- β receptors, and the effects of the Type I receptor on binding of TGF- β to the Type II receptor.

Type I receptor proteins have been reported not to bind to their ligand molecules independently, but, acting in concert with Type II receptor proteins, are observed to contribute to increased binding to the ligand. See Matsuzaki et al., <u>J. Biol. Chem.</u>, 268:12719-12723 (1993); Ebner et al., <u>Science</u>, 260:1344-1348 (1993).

Paralkar et al., <u>PNAS USA</u> <u>88</u>:3397-3401 (1991) describes the presence of high affinity binding sites for BMP-4 on MC3T3E1 and NIH3T3 cells. No competition by TGF- β was found for the BMP-4 binding proteins, nor was competition by BMP-4 for TGF- β receptors observed in Attisano et al., <u>Cell</u> <u>68</u>:97-108 (1992).

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SUMMARY OF THE INVENTION

In one embodiment, the present invention comprises a purified and isolated DNA molecule which encodes a BMP receptor protein, said DNA molecule preferably comprising the clones CFK1-43a and CFK1-23a, or a DNA sequence selected from the group consisting of SEQ ID NO:1 and SEQ ID NO:3.

The present invention further comprises purified and isolated DNA molecules which encode BMP receptor proteins, said BMP receptor proteins preferably comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2 and SEQ ID NO:4. In another embodiment, the present invention comprises a BMP receptor protein CFK1-43a and CFK1-23a, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2 and SEQ ID NO:4.

The present invention further comprises DNA molecules comprising a DNA sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7 and SEQ ID NO:9, and DNA molecules which encode serine/threonine kinase receptor proteins comprising an amino acid sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8 and SEQ ID NO:10. These DNA molecules and proteins are related to the BMP family of receptors. Among other uses, these DNA molecules are presently useful as probes for isolating and purifying additional novel BMP receptors.

The present invention also comprises novel DNA sequences which encode receptor proteins, which novel DNA sequences are identified by a method using DNA sequence encoding all or a fragment of the receptor proteins of the present invention. In preferred embodiments, the novel DNA sequences are identified using DNA sequence from the serine/threonine kinase domain of a receptor, which is highly conserved among the family of BMP receptors. Alternatively, DNA sequence encoding the ligand binding domain could be used to identify additional novel BMP receptor encoding sequences.

The present invention further comprises DNA molecules encoding soluble, truncated receptor proteins, and the soluble proteins themselves. The truncated receptor proteins preferably comprise the ligand binding domain, but not the serine/threonine kinase and transmembrane domains, of the receptor protein. The truncated receptor proteins are soluble, and will be secreted into supernatant by mammalian cells. Thus, when expressed

in mammalian cells using a DNA molecule encoding a truncated receptor protein, the truncated receptor protein will be secreted rather than expressed on the surface of the host cell. The truncated receptor protein thereby expressed still binds specifically to BMPs, and can be used to block receptors from mediating the cellular processes in which they normally participate in as signalling mechanisms by competition for the same ligand. The truncated receptor protein could compete with receptor proteins normally expressed on the surface of responsive cells for functional ligand and inhibit the formation of a functional receptor-ligand complex, thereby blocking the normal signalling mechanism of the complex and the cellular processes normally affected by functional receptor-ligand interactions.

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In one aspect, the invention provides a method for producing cells expressing more than one receptor protein comprising culturing a selected host cell containing a polynucleotide sequence encoding a first selected receptor protein, truncated receptor protein, or active fragment thereof and a polynucleotide sequence encoding a second selected receptor protein, truncated receptor protein, or active fragment thereof. The resulting cells, which will express multiple co-expressed, biologically active receptors, may be isolated and used in a therapeutic composition.

Another aspect of the current invention comprises ligands for the BMP receptors and truncated BMP receptor protein, said ligands being characterized by the ability to bind to the receptors. Such ligands may stimulate growth of bone and/or cartilage, or may be involved in influencing other developmental processes. Said ligands may be monoclonal antibodies, small peptide BMP analogues, or small organic molecule BMP analogues as further characterized herein. In a preferred embodiment, said ligands comprise antibodies against the truncated, soluble receptor protein and the receptor proteins of the invention. These antibodies can be employed in a variety of diagnostic and therapeutic applications. Such antibodies can be used to identify cell types which naturally express receptors of the invention and may therefore have the capacity to elicit a biological response upon exposure to the appropriate ligand. These antibodies can be further useful in the identification of additional receptor proteins capable of binding to other individual BMPs and/or BMP heterodimers. Additionally such antibodies are useful in blocking the formation of functional receptor-ligand complexes and thus inhibit the cellular responses that would

normally be mediated by these complexes. Alternatively, such antibodies may mimic the effect of BMP by interacting with the receptor in a way that would stimulate the cellular responses that would normally be mediated by a functional receptor-ligand complex.

In yet another embodiment, the invention comprises pharmaceutical compositions comprising a compound first identified for such use as a ligand for the truncated BMP receptor and therapeutic methods for the treatment of bone and/or cartilage disorders comprising administering a ligand for the truncated BMP receptor.

Other aspects and advantages of the present invention will be apparent upon consideration of the following detailed description and preferred embodiments thereof.

Brief Description of the Sequences

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SEQ ID NO:1 comprises DNA and amino acid sequence of the BMP receptor protein CFK1-23a, isolated from rat cell line CFK1. This DNA contained in plasmid CFK1-23a, which has been deposited and accorded ATCC #69378, further described below.

SEQ ID NO:2 comprises the amino acid sequence encoded by the CFK1-23a DNA sequence.

SEQ ID NO:3 comprises DNA and amino acid sequence of the BMP receptor protein CFK1-43a, isolated from rat cell line CFK1. This DNA contained in plasmid CFK1-43a has been deposited and accorded ATCC #69381, further described below.

SEQ ID NO:4 comprises the amino acid sequence encoded by the CFK1-43a DNA sequence.

SEQ ID NO:5 comprises DNA and amino acid sequence of the serine/threonine kinase receptor protein CFK1-10a, isolated from rat cell line CFK1. This DNA contained within plasmid CFK1-10a has been deposited and accorded ATCC #69380, further described below.

SEQ ID NO:6 comprises the amino acid sequence encoded by the CFK1-10a DNA sequence.

SEQ ID NO:7 comprises DNA and amino acid sequence of the serine/kinase receptor protein W101, isolated from murine cell line W-20-17. This DNA contained in plasmid pMT101 has been deposited and accorded ATCC #69379, further described

below.

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SEQ ID NO:8 comprises the amino acid sequence encoded by the W101 DNA sequence.

SEQ ID NO:9 comprises DNA and amino acid sequence of the serine/kinase receptor protein W120, isolated from murine cell line W-20-17. This DNA contained in plasmid pMT120E has been deposited and accorded ATCC #69377, further described below.

SEQ ID NO:10 comprises the amino acid sequence encoded by the W120 DNA sequence.

SEQ ID NO:11 comprises DNA and amino acid sequence of the serine/kinase receptor protein KDA-B5. This DNA was used as a probe to identify novel serine/kinase receptors of the present invention.

SEQ ID NO:12 comprises the amino acid sequence encoded by the KDA-B5 DNA sequence.

SEQ ID NO:13: comprises the DNA sequence of oligonucleotide primer A.

SEQ ID NO:14: comprises the DNA sequence of oligonucleotide primer B.

SEQ ID NO:15: comprises the DNA sequence of oligonucleotide primer C..

SEQ ID NO:16 comprises the DNA sequence of oligonucleotide primer D.

SEQ ID NO:17 comprises the DNA sequence of oligonucleotide primer E.

SEQ ID NO:18: comprises the amino acid sequence of a portion of KDA-B5 used to design oligonucleotide primer A.

SEQ ID NO:19 comprises the amino acid sequence of a portion of KDA-B5 used to design oligonucleotide primer B through E.

Detailed Description of the Invention

Bone morphogenetic proteins are characterized by their ability to promote, stimulate or otherwise induce the formation of cartilage and/or bone. The ability of these proteins to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below. These proteins can be used in compositions which may be used to induce bone and/or cartilage formation. These BMP compositions may also be used for wound healing and tissue repair. Further uses of such compositions include the treatment

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of bone and/or cartilage defects, periodontal disease and other tooth repair processes, treatment of osteoporosis and increase of neuronal survival.

The BMP receptors and truncated receptors of the present invention are useful, among other uses, for the identification of BMPs, the identification of further BMP receptors, and the identification of ligands or molecules, including antibodies, which are able to mimic the binding characteristics of BMPs. These ligands may act as agonist or antagonists, depending upon the individual ligand. The activity of the ligands may be characterized in an assay for BMP activity, such as the W-20-17 alkaline phophatase induction assay and rat ectopic bone formation assay, described at Examples XII and XIII below. The BMP receptors are also useful in inhibiting the effects of BMPs, where such inhibition is desired.

BMP receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-532 of SEQ ID NO:2; or amino acid # 1-502 of SEQ ID NO:4.

The purified human BMP receptor proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:1 from nucleotide # 61 to nucleotide 1656 (or to 1659 with the stop codon); or SEQ ID NO:3 from nucleotide #247 to nucleotide 1752 (or to 1755 stop codon); and recovering and purifying from the transformed cell membrane a protein which contains the derived amino acid sequence, or a substantially homologous sequence as represented by amino acid # 24 to # 532 of SEQ ID NO:2; or amino acid # 8 to # 502 of SEQ ID NO:4. Since the BMP receptor proteins expressed in this manner are expected to remain associated with the cell membrane of the transformed cell, recombinant receptor proteins of the invention can be dissociated from the transformed cell membrane and are then purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

Truncated BMP receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-149 of SEQ ID NO:2; or amino acid # 1-124 of SEQ ID NO:4.

The purified human truncated BMP receptor proteins of the present invention may

be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:1 from nucleotide # 61 to nucleotide 507; or SEQ ID NO:3 from nucleotide # 247 to nucleotide 618; and recovering and purifying from the culture medium a protein which contains the derived amino acid sequence, or a substantially homologous sequence, as represented by amino acid # 24 to # 149 of SEQ ID NO:2; or amino acid # 8 to # 124 of SEQ ID NO:4. In the above amino acid sequences, the secretory leader sequence (e.g., amino acids 1 to 23 of SEQ ID NO:2) will not be present since these are typically cleaved away from secreted proteins. The leader sequence predicted for SEQ ID NO:4 by standard computer programs is amino acids 1 to 7; however, it is contemplated that the actual leader sequence may be longer since seven amino acids is unusually short for a leader sequence. Thus, the protein purified from culturing host cells transformed with a DNA molecule comprising the DNA sequence of SEQ ID NO:3 from nucleotide # 247 to nucleotide 618 may be shorter than amino acid # 8 to # 124 of SEQ ID NO:4.

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The truncated BMP receptor proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

Other serine/threonine kinase receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-509 of SEQ ID NO:6.

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The purified serine/threonine kinase receptor proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:5 from nucleotide # 474 to nucleotide 2000 (or to 2003 stop codon); and recovering and purifying from the transformed cell membrane a protein which contains the derived amino acid sequence, or a substantially homologous sequence as represented by amino acid # 18 to # 509 of SEQ ID NO:6. Since the serine/threonine kinase receptor proteins expressed in this manner are expected to remain associated with the cell membrane of the transformed cell, recombinant receptor proteins of the invention can be dissociated from the transformed cell membrane and are then purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

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Truncated serine/threonine kinase receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-121 of SEQ ID NO:6.

The purified human truncated serine/threonine kinase receptor proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:5 from nucleotide # 474 to nucleotide 836 and recovering and purifying from the culture medium a protein which contains the derived amino acid sequence, or a substantially homologous sequence as represented by amino acid # 18 to # 121 of SEQ ID NO:6. The truncated serine/threonine kinase receptor proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

Serine/kinase receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-505 of SEQ ID NO:8; or amino acid # 1-503 of SEQ ID NO:10.

The purified serine/kinase receptor proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:7 from nucleotide # 80 to nucleotide 1594 (or to 1597 stop codon); or SEQ ID NO:9 from nucleotide # 83 to nucleotide 1591 (or to 1594 stop codon); and recovering and purifying from the transformed cell membrane a protein which contains the derived amino acid sequence, or a substantially homologous sequence as represented by amino acid # 24 to # 505 of SEQ ID NO:8; or amino acid # 30 to # 503 of SEQ ID NO:10. Since the serine/kinase receptor proteins expressed in this manner are expected to remain associated with the cell membrane of the transformed cell, recombinant receptor proteins of the invention can be dissociated from the trans formed cell membrane and are then purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

Truncated serine/threonine kinase receptor proteins of the present invention may be characterized by an amino acid sequence comprising amino acid # 1-122 of SEQ ID NO:8; or amino acid # 1-121 of SEQ ID NO:10.

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in the invention.

The purified human truncated serine/threonine kinase receptor proteins of the present invention may be produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of SEQ ID NO:7 from nucleotide # 80 to nucleotide 445; or SEQ ID NO:9 from nucleotide # 83 to nucleotide 445; and recovering and purifying from the culture medium a protein which contains the derived amino acid sequence, or a substantially homologous sequence, as represented by amino acid # 24 to # 122 of SEQ ID NO:8; or amino acid # 30 to # 121 of SEQ ID NO:10. The truncated serine/threonine kinase receptor proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials with which they are co-produced and from other contaminants present.

The present invention also encompasses DNA molecules comprising the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding for the expression of the above receptor proteins. These DNA sequences include those depicted in SEQ ID NOS: 1, 3, 5, 7 and 9, in a 5' to 3' direction and those sequences which hybridize under stringent hybridization conditions [see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] to the DNA sequences of SEQ ID NOS: 1, 3, 5, 7 and 9; and encode a protein having the ability to bind to BMP or which is useful to isolate novel BMP receptors.

Similarly, DNA sequences which code for the above receptor polypeptides coded for by the amino acid sequences of SEQ ID NO: 2, 4, 6, 8 and 10, but which differ in codon sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the novel receptor proteins described herein. Variations in the DNA sequences of SEQ ID NOS: 1, 3, 5, 7 and 9 which are caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded thereby are also encompassed

Another aspect of the present invention provides a novel method for producing receptor proteins. The method of the present invention involves culturing a suitable cell

line, which has been transformed with a DNA molecule comprising a DNA sequence coding on expression for a receptor protein, under the control of known regulatory sequences. The transformed host cells are cultured and the receptor proteins recovered and purified from the transformed cell membrane. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Another aspect of the present invention provides a novel method for producing truncated receptor proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA molecule comprising a DNA sequence coding on expression for a truncated receptor protein, under the control of known regulatory sequences. The transformed host cells are cultured and the truncated receptor proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

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Suitable cells or cell lines for production of the receptor proteins or truncated receptor proteins may be mammalian cells, such as Chinese hamster ovary cells (CHO) or BHK cells. The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell line CV-1 may also be suitable.

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Bacterial cells may also be suitable hosts. For example, the various strains of <u>E. coli</u> (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of <u>B. subtilis</u>, <u>Pseudomonas</u>, other bacilli and the like may also be employed in this method.

Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present

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invention. See, e.g. Miller et al, <u>Genetic Engineering</u>, <u>8</u>:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in expression of these novel receptor polypeptides. Preferably, the vectors contain the full novel DNA sequences described above which encode the novel receptor proteins of the invention. Additionally, the vectors contain appropriate expression control sequences permitting expression of the receptor protein sequences.

Alternatively, vectors incorporating modified DNA sequences as described above are also embodiments of the present invention and useful in the production of the receptor proteins. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Useful regulatory sequences for such vectors are known to one of skill in the art and may be selected depending upon the selected host cells. Such selection is routine and does not form part of the present invention.

The BMP receptor proteins of the present invention, such as CFK1-23a and CFK1-43a, have been found to bind to members of the BMP family, preferably BMP-2 and BMP-4, but not to TGF- β . Thus, the BMP receptor proteins of the present invention are distinguished from TGF- β receptors, which bind to TGF- β .

The present invention may include co-transfection of cells with DNA molecules comprising DNA sequences encoding multiple receptor proteins in order to achieve binding to a ligand molecule such as a BMP. Thus, for example, a DNA molecule comprising a DNA sequence encoding the receptor protein CFK1-10a may be co-transfected into cells along with a DNA molecule comprising a DNA sequence encoding receptor protein CFK1-23a or CFK1-43a.

The DNA molecules comprising DNA sequences encoding the receptor proteins of the present invention are useful for the production of cells which express receptor proteins. These cells, when transformed with the DNA molecules of the present invention, will express receptor proteins on their surface. In turn, these cells will bind more readily to the ligand and may demonstrate increased responsiveness to the ligand. For example, cells

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which express the BMP receptor proteins of the present invention exhibit increased binding to BMP-2 and BMP-4, and will exhibit increased responsiveness to BMPs such as BMP-2 and BMP-4. The increased BMP response is desirable for accelerating the effects of BMPs, which include the osteoinductive promotion of bone growth and cartilage regeneration.

The BMP receptor proteins of the present invention are useful for isolating BMP. Additionally, BMP receptor proteins of the invention are useful in the identification of novel molecules related to BMPs which may be capable of inducing the formation of bone or cartilage or may be involved in influencing other developmental processes. In addition, the BMP receptor proteins are useful for identifying and/or quantifying BMP-2 and/or BMP-4 in a sample, as well as for inhibiting the effects of BMP-2 or BMP-4 on cells. The BMP receptors of the present invention may further be useful in identifying synthetic and naturally-occurring chemical entities which are able to mimic the binding effects of BMP-2 and/or BMP-4. The BMP receptor proteins of the invention may also be useful in identifying synthetic and naturally-occurring chemical entities which are able to antagonize and/or inhibit the binding effects of BMP-2 and/or BMP-4. The BMP receptor proteins may also be useful in identifying compounds which play a role in regulating the expression of BMP receptor proteins. Those compounds could be used in order to stimulate BMP-responsiveness, for example, bone growth, in particular tissues or cells of interest.

The novel serine/threonine kinase receptor proteins of the present invention also include W101 and W120, which have been isolated from murine cell line W-20-17, a cell line which is known to be responsive to BMP. The DNA encoding one of these novel receptor proteins has been used as a probe in order to isolate other clones which are potentially members of the class of BMP receptor proteins, including the CFK1-23a and CFK1-43a clones, which have been confirmed to encode proteins which are members of the BMP receptor family. Thus, the DNA molecules comprising DNA sequence encoding the serine/kinase receptor proteins of the present invention are useful for the isolation of DNA encoding BMP receptor proteins, and the present invention includes such a method of using the DNA molecules comprising DNA sequence encoding serine/threonine kinase

receptor proteins, as well as the novel BMP receptor proteins which are thereby isolated.

In one embodiment of the present invention, novel DNA sequences which encode BMP receptor proteins are identified by a method using DNA sequence encoding all or a fragment of the serine/kinase receptor proteins of the present invention. In preferred embodiments, the novel DNA sequences are identified using DNA sequence encoding the serine/threonine kinase domain of a receptor. Alternatively, DNA sequence encoding the ligand binding domain could be used to identify additional novel BMP receptor encoding sequences.

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Thus, the present invention further comprises methods of identifying new BMP receptor proteins and DNA molecules encoding those proteins, and the proteins and DNA molecules thus identified. The method comprises preparing a DNA fragment which encodes a selected domain of a BMP receptor protein, preferably the kinase domain of a BMP receptor protein, or alternatively a DNA fragment encoding the ligand binding domain, and using that fragment as a probe to screen either a genomic or cDNA library. The cDNA library is preferably prepared from a cell line known to express BMP receptors. These include the murine cell line W-20-17 and the rat cell line CFK1. The DNA sequences which are thus identified share homology with the known BMP receptor protein, and thus are expected to encode a protein which will bind to one or more BMPs. Using methods known in the art, one can clone the entire DNA sequence which is thereby identified and use it to express the newly identified BMP receptor protein. Identification of the new protein as a BMP receptor protein is confirmed using the binding assay described in Example VI.

Another embodiment of the present invention comprises DNA molecules comprising DNA sequences encoding truncated receptor proteins, and the truncated proteins themselves. The truncated receptor proteins preferably comprise the ligand binding domain, but not the serine/threonine kinase and transmembrane domains, of the receptor protein. The truncated receptor proteins are soluble, and will be secreted into supernatant by mammalian cells. Thus, when expressed in mammalian cells using a DNA molecule encoding a truncated receptor protein, the truncated receptor protein will be secreted rather than expressed on the surface of the host cell. The truncated receptor protein thereby

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expressed still binds specifically to its ligand. Thus, the truncated BMP receptor proteins can be used to block BMP receptors of the invention from mediating the cellular processes in which they normally participate in as signalling mechanisms. The truncated receptor protein could compete with receptor proteins normally expressed on the surface of responsive cells for functional ligand and inhibit the formation of a functional receptor-ligand complex, thereby blocking the normal signalling mechanism of the complex and the cellular processes normally affected by functional receptor-ligand interactions.

Compositions containing the truncated BMP receptor proteins of the present invention may be used for the inhibition of the effects of BMPs such as BMP-2 and/or BMP-4 on cells. The present invention includes therapeutic methods comprising administering such a composition topically, systematically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the desired site. Therapeutically useful agents, such as growth factors (e.g., BMPs, $TGF-\beta$, FGF, IGF), cytokines (e.g., interleukins and CSFs) and antibiotics, may also optionally be included in or administered simultaneously or sequentially with, the receptor composition in the methods of the invention.

Another embodiment of the present invention comprises cells which have been transformed with the DNA molecules comprising DNA sequences encoding the BMP receptor proteins of the present invention. These cells will express BMP receptors on their surface, which will increase the cells' responsiveness to BMP. Thus, cells transformed with the DNA molecules encoding the BMP receptor proteins may be administered therapeutically, to promote response to BMP, for example, bone and/or cartilage regeneration at a desired site.

There is a wide range of methods which can be used to deliver the cells expressing BMP receptor proteins to a site for use in promoting a BMP response such as bone and or cartilage regeneration. In one embodiment of the invention, the cells expressing BMP receptor protein can be delivered by direct application, for example, direct injection of a sample of such cells into the site of bone or cartilage damage. In a particular embodiment,

these cells can be purified. In a preferred embodiment, the cells expressing BMP receptor protein can be delivered in a medium or matrix which partially impedes their mobility so as to localize the cells to a site of bone or cartilage injury. Such a medium or matrix could be semi-solid, such as a paste or gel, including a gel-like polymer. Alternatively, the medium or matrix could be in the form of a solid, preferably, a porous solid which will allow the migration of cells into the solid matrix, and hold them there while allowing proliferation of the cells.

Expression of Receptor Protein

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In order to produce receptor protein, the DNA encoding the desired protein is transferred into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The presently preferred expression system for biologically active recombinant receptor protein is stably transformed mammalian cells.

One skilled in the art can construct mammalian expression vectors by employing the sequence of SEQ ID NO: 1, 3, 5, 7 or 9, or other DNA sequences containing the coding sequences of SEQ ID NO: 1, 3, 5, 7 or 9, or other modified sequences and known

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vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)] and pJL3, pJL4 [Gough et al., EMBO I., 4:645-653 (1985)]. The receptor protein cDNA sequences can be modified by removing the non-coding nucleotides adjacent to the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. The transformation of these vectors into appropriate host cells can result in expression of receptor proteins.

One skilled in the art can manipulate the sequences of SEQ ID NO: 1, 3, 5, 7 or 9 by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells. For example, the coding sequences can be further manipulated (e.g. ligated to other known linkers or modified by deleting noncoding sequences therefrom or altering nucleotides therein by other known techniques). The modified receptor protein coding sequence can then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci.USA, 77:5230-5233 (1980). This exemplary bacterial vector can then be transformed into bacterial host cells and receptor protein expressed thereby. For a strategy for producing extracellular expression of receptor proteins in bacterial cells., see, e.g. European patent application EPA 177,343.

Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector can also be constructed employing yeast regulatory sequences for intracellular or extracellular expression of the receptor proteins of the present invention by yeast cells. [See, e.g., procedures described in published PCT application WO86/00639 and European patent application EPA 123,289].

A method for producing high levels of a receptor protein of the invention in mammalian cells involves the construction of cells containing multiple copies of one or more of the heterologous receptor genes. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp,

J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

For example, a plasmid containing a DNA sequence for a BMP receptor protein of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by calcium phosphate coprecipitation and transfection, electroporation, protoplast fusion or lipofection. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., 5:1750 (1983).

Transformants are cloned, and binding to BMP-2 or BMP-4 is measured by the binding assay described above in Example VI. BMP-2 and BMP-4 binding should increase with increasing levels of MTX resistance. Similar procedures can be followed to produce other related BMP receptor proteins.

Co-Expression of Multiple Receptor Proteins

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According to one embodiment of this invention, the host cell may be co-transfected with one or more vectors containing coding sequences for one or more receptor proteins, truncated receptor proteins or active fragments thereof. Each receptor polynucleotide sequence may be present on the same vector or on individual vectors co-transfected into the cell. Alternatively, the polynucleotides encoding receptors, truncated receptors or their fragments may be incorporated into a chromosome of the host cell. Additionally, a single transcription unit may encode single copy of two genes encoding different receptor proteins.

According to another embodiment of this invention, the selected host cell containing the two polypeptide encoding sequences is a hybrid cell line obtained by fusing two selected, stable host cells, each host cell transfected with, and capable of stably expressing, a polynucleotide sequence encoding a selected first or second receptor protein, truncated receptor protein or active fragment thereof.

In another aspect of the present invention, therefore, there are provided

compositions of cells which express more than one recombinant receptor protein, truncated receptor protein, or active fragments thereof which retain the binding characteristics of the receptor or truncated receptor. Also provided are compositions of truncated truncated receptor proteins secreted by host cells. The cells, proteins, and compositions of receptor proteins, truncated receptor proteins or active fragments thereof may be characterized by their ability to bind selectively to BMPs with greater binding affinity than to other proteins in the TGF- β superfamily in a binding assay.

The cells and compositions may comprise one or more BMP receptor proteins, truncated BMP receptor proteins, or active fragments thereof; or of one or more serine/threonine kinase receptor proteins, truncated serine/threonine kinase receptor proteins, or active fragments thereof, such as W-101, W-120 or CFK1-10a, in combination with one or more BMP receptor proteins, truncated BMP receptor proteins, or active fragments thereof, such as CFK1-23a or CFK1-43a. These cells or compositions may be produced by co-expressing each protein in a selected host cell and isolating the cells in a composition or, in the case where truncated receptor proteins are produced, by isolating the truncated receptor proteins from the culture medium.

As a further aspect of this invention a cell line is provided which comprises a first polynucleotide sequence encoding a first receptor protein, truncated receptor protein, or active fragment thereof and a second polynucleotide sequence encoding a second receptor protein, truncated receptor protein, or active fragment thereof, the sequences being under control of one or more suitable expression regulatory systems capable of co-expressing the receptor proteins. The cell line may be transfected with one or more than one polynucleotide molecule. Alternatively, the cell line may be a hybrid cell line created by cell fusion as described above.

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Another aspect of the invention is a polynucleotide molecule or plasmid vector comprising a polynucleotide sequence encoding a first selected receptor protein, truncated receptor protein, or active fragment thereof and a polynucleotide sequence encoding a second selected receptor protein, truncated receptor protein, or active fragment thereof. The sequences are under the control of at least one suitable regulatory sequence capable of directing co-expression of each protein or active fragment. The molecule may contain

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a single transcription unit containing a copy of both genes, or more than one transcription unit, each containing a copy of a single gene.

One embodiment of the method of the present invention for producing compositions of cells or recombinant receptor proteins involves culturing a suitable cell line, which has been co-transfected with a DNA sequence coding for expression of a first receptor protein, truncated receptor protein, or active fragment thereof and a DNA sequence coding for expression of a second receptor protein, truncated receptor protein, or active fragment thereof, under the control of known regulatory sequences. The transformed host cells are cultured and the cells are isolated and purified to form compositions of transformed cells. In the embodiment wherein truncated receptor proteins are produced, the truncated receptor protein is recovered and purified from the culture medium and can be used to form compositions of truncated receptor protein.

In another embodiment of this method which is the presently preferred method of expression of the recombinant receptor proteins of this invention, a single host cell, e.g., a CHO DUKX cell, is co-transfected with a first DNA molecule containing a DNA sequence encoding one receptor protein, such as the receptor protein CFK1-10a, and a second DNA molecule containing a DNA sequence encoding a second selected receptor protein, such as the BMP receptor protein CFK1-23a or CFK1-43a. One or both plasmids contain a selectable marker that can be used to establish stable cell lines expressing the receptor proteins. These separate plasmids containing distinct receptor genes on separate transcription units are mixed and transfected into the CHO cells using conventional protocols. A ratio of plasmids that gives maximal expression of activity in the binding assay can be determined.

For example, equal ratios of a plasmid containing the first receptor protein gene and a dihydrofolate reductase (DHFR) marker gene and another plasmid containing a second receptor protein gene and a DHFR marker gene can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by calcium phosphate coprecipitation and transfection, electroporation, microinjection, protoplast fusion or lipofection. Individual DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum by conventional means. DHFR+ cells containing increased gene copies can be

selected for propagation in increasing concentrations of methotrexate (MTX) (e.g. sequential steps in 0.02, 0.1, 0.5 and 2.0 uM MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982); and Kaufman et al, Mol. Cell Biol., 5:1750 (1983). Expression of or at least one receptor protein linked to DHFR should increase with increasing levels of MTX resistance. Cells that stably express either or both receptor protein/DHFR genes will survive. However at a high frequency, cell lines stably incorporate and express both plasmids that were present during the initial transfection. The conditioned medium is thereafter harvested and the receptor protein isolated by conventional methods and assayed for activity. This approach can be employed with DHFR-deficient cells.

As an alternative embodiment of this method, a DNA molecule containing one selected receptor gene may be transfected into a stable cell line which already expresses another selected receptor gene. For example, a stable CHO cell line expressing the gene for receptor CFK1-10a with the DHFR marker may be transfected with a plasmid containing the gene for receptor gene for CFK1-23a and a second selectable marker gene, e.g., neomycin resistance (Neo). After transfection, the cell is cultured and suitable cells selected by treatment with MTX and the antibiotic, G-418. Surviving cells are then screened for the expression of both receptor proteins. This expression system has the advantage of permitting a single step selection.

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Alternative dual selection strategies using different cell lines or different markers can also be used. For example, the use of an adenosine deaminase (ADA) marker to amplify the second receptor gene in a stable CHO cell line expressing a different receptor with the DHFR marker may be preferable, since the level of expression can be increased using deoxycoformycin (DCF)-mediated gene amplification. Alternatively, any cell line expressing a receptor made by first using this marker can then be the recipient of a second receptor expression vector containing a distinct marker and selected for dual resistance and receptor coexpression.

Still another embodiment of a method of expressing the receptors of this invention includes transfecting the host cell with a single DNA molecule encoding multiple genes for expression either on a single transcription unit or on separate transcription units.

Multicistronic expression involves multiple polypeptides encoded within a single transcript, which can be efficiently translated from vectors utilizing a leader sequence, e.g., from the EMC virus, from poliovirus, or from other conventional sources of leader sequences. Two receptor genes (Rx and Ry, respectively) and a selectable marker can be expressed within a single transcription unit. For example, vectors containing the configuration Rx-EMC-Ry-DHFR or Rx-EMC-Ry-EMC-DHFR can be transfected into CHO cells and selected and amplified using the DHFR marker. A plasmid may be constructed which contains DNA sequences encoding two different receptors, one or more marker genes and a suitable leader or regulatory sequence on a single transcription unit.

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Similarly, host cells may be transfected with a single plasmid which contains separate transcription units for each receptor. A selectable marker, e.g., DHFR, can be contained on a another transcription unit, or alternatively as the second cistron on one or both of the receptor genes. These plasmids may be transfected into a selected host cell for expression of the receptors.

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Another embodiment of this expression method involves cell fusion. Two stable cell lines which express selected receptors, such as a cell line transformed with a vector for CFK1-23a (e.g., pMV23a) and a cell line stably transformed with a vector for CFK1-43a (e.g., pMV43a), developed using the DHFR/MTX gene amplification system and expressing receptors at high levels, can be transfected with one of several dominant marker genes (e.g., neo^r, hygromycin^r, GPT). After sufficient time in coculture (approximately one day) one resultant cell line expressing one receptor and a dominant marker can be fused with a cell line expressing a different receptor and preferably a different marker using a fusigenic reagent, such as polyethylene glycol, Sendai virus or other known agent.

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The resulting cell hybrids expressing both dominant markers and DHFR can be selected using the appropriate culture conditions, and screened for coexpression of the receptors, truncated receptors or their fragments. The selected hybrid cell contains sequences encoding both selected receptors, and both receptors will be retained within the membrane of the cell. Compositions of the cells expressing multiple receptors can be used in the methods of the present invention to interact with BMP. In the case where genes encoding truncated receptors are used, the truncated receptor is formed in the cell and then

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secreted. The truncated receptor protein is obtained from the conditioned medium and isolated and purified therefrom by conventional methods. The resulting receptor protein composition may be characterized by methods described herein and may be used in the methods of the present invention, for example, to compete with receptors present in cells for ligand binding and thus inhibit the activity of BMP.

Cell lines generated from the approaches described above can be used to produce co-expressed receptor polypeptides. The receptor proteins are retained within the membrane of the cells. Compositions of the cells may be used in order to increase response to BMP, for example to increase cartilage and/or bone formation. Compositions of the cells may be applied in conjunction with BMP.

Where truncated receptor polypeptides are produced, the receptor proteins are isolated from the cell medium in a form substantially free from other proteins with which they are co-produced as well as from other contaminants found in the host cells by conventional purification techniques. The presently preferred method of production is co-transfection of different vectors into CHO cells and methotrexate-mediated gene amplification. Stable cell lines may be used to generate conditioned media containing truncated receptor that can be purified and assayed for in vitro and in vivo activities. For example, the resulting truncated receptor-producing cell lines obtained by any of the methods described herein may be screened for activity by the binding assays described in Example VI, RNA expression, and protein expression by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE).

The above-described methods of co-expression of the receptors of this invention utilize suitable host cells or cell lines. Suitable cells preferably include mammalian cells, such as Chinese hamster ovary cells (CHO). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U. S. Patent 4,419,446. Other suitable mammalian cell lines are the CV-1 cell line, BHK cell lines and the 293 cell line.

Another aspect of the present invention provides DNA molecules or plasmid vectors

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for use in expression of these recombinant receptor proteins. These plasmid vectors may be constructed by resort to known methods and available components known to those of skill in the art. In general, to generate a vector useful in the methods of this invention, the DNA encoding the desired receptor protein, truncated receptor protein, or active fragment thereof, is transferred into one or more appropriate expression vectors suitable for the selected host cell.

It is presently contemplated that any expression vector suitable for efficient expression in mammalian cells may be employed to produce the recombinant receptor proteins of this invention in mammalian host cells. Preferably the vectors contain the selected receptor DNA sequences described above and in the Sequence Listings, which encode selected receptor proteins, or truncated receptor proteins. Alternatively, vectors incorporating modified sequences are also embodiments of the present invention and useful in the production of the vectors.

In addition to the specific vectors described above, one skilled in the art can construct mammalian expression vectors by employing the sequence of SEQ ID NO:1,3,5,7, or 9 or other DNA sequences coding for receptor proteins, truncated receptor proteins, or active fragments thereof and known vectors, such as pCD [Okayama et al, Mol. Cell Biol., 2:161-170 (1982)] and pJL3, pJL4 [Gough et al, EMBO J., 4:645-653 (1985)]. The receptor DNA sequences can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. The transformation of these vectors into appropriate host cells as described above can produce desired receptor proteins.

One skilled in the art could manipulate the sequences of SEQ ID NO:1,3,5,7, or 9 by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with e.g., yeast or insect regulatory sequences, to create vectors for intracellular or extracellular expression by yeast or insect cells. [See, e.g., procedures described in published European Patent Application 155,476] for expression in insect cells; and procedures described in published PCT application WO86/00639 and European Patent Application EPA 123,289 for expression in yeast cells].

Similarly, bacterial sequences and preference codons may replace sequences in the described and exemplified mammalian vectors to create suitable expression systems for use in the production of receptor proteins in the method described above. For example, the coding sequences could be further manipulated (e.g., ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified receptor coding sequences could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al, Proc. Natl. Acad. Sci. USA, 77:5230-5233 (1980). The exemplary bacterial vector could then be transformed into bacterial host cells and receptor proteins expressed thereby.

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Other vectors useful in the methods of this invention may contain multiple genes in a single transcription unit. For example, a proposed plasmid contains the CFK1-10a receptor gene followed by the EMC leader sequence, followed by the CFK1-23a BMP receptor gene, followed by the DHFR marker gene. Another example contains the CFK1-23a BMP receptor gene, the EMC leader, the W101 serine/threonine kinase receptor gene, another EMC leader sequence and the DHFR marker gene. Alternatively, the vector may contain more than one transcription unit. As one example, the plasmid may contain a transcription unit for CFK1-23a BMP receptor gene and a separate transcription unit for CFK1-43a receptor gene, i.e., CFK1-23a-EMC-DHFR and CFK1-43a-EMC-DHFR. Alternatively, each transcription unit on the plasmid may contain a different marker gene. For example, the plasmid may contain CFK1-10a-EMC-Neo and CFK1-43a-EMC-DHFR. Of course, the above examples are not limiting. Other combinations (i.e., co-expression) of the receptors of the present invention are also within the invention.

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Additionally the vectors also contain appropriate expression control sequences which are capable of directing the replication and expression of the receptor in the selected host cells. Useful regulatory sequences for such vectors are known to one of skill in the art and may be selected depending upon the selected host cells. Such selection is routine and does not form part of the present invention. Similarly, the vectors may contain one or more selection markers, such as the antibiotic resistance gene, Neo or selectable markers such as DHFR and ADA. The presently preferred marker gene is DHFR. These marker genes may also be selected by one of skill in the art.

The following examples illustrate practice of the present invention in recovering and characterizing the receptor proteins of the present invention and employing them to recover the corresponding human receptor proteins of the present invention, and in expressing the proteins via recombinant techniques.

5 Antibodies

As used herein, a molecule is said to be "BMP-like" if it exhibits at least one BMP-like activity. For the purposes of the invention, BMP-like activity includes the ability to bind to a truncated BMP receptor and to stimulate growth of BMP-dependent cell lines such as the W-20-17 cell line described in Example VIII. As used herein, the term "BMP-like monoclonal antibody" includes non-human BMP-like monoclonal antibodies, complementarity determining regions (CDRs) of the non-human BMP-like monoclonal antibodies, BMP recognition sites of the non-human BMP-like monoclonal antibodies, all engrafted forms of the BMP recognition sites of the non-human BMP-like monoclonal antibodies, and small peptide BMP analogues.

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In accordance with the present invention, the term "BMP analogue" encompasses monoclonals, small peptides, and small molecules which possess BMP-like activity. For the purposes of the invention, BMP-like activity is defined as the ability to bind to the truncated BMP receptor and to stimulate growth of BMP-dependent cell lines such as the W-20-17 cell line described in Example VIII.

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The BMP-like monoclonal antibodies of the invention may be molecularly altered to enhance their utility as human pharmaceuticals. For example, the CDRs of the BMP-like monoclonals comprise specific BMP receptor recognition sites, which are encompassed in the present invention. These CDRs may be engrafted onto human immunoglobulin framework regions to minimize antigenicity caused by the presence of non-human immunoglobulin regions, for example by the techniques disclosed in WO 91/09967. The BMP receptor recognition sites of the present invention may also be engrafted onto other protein frameworks to maximize the therapeutic efficiency of the BMP analogues developed therefrom. The CDRs of the BMP-like monoclonals may also be molecularly altered to form single chain antibodies.

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The CDRs of the BMP-like monoclonal antibodies may also be used to make small

peptide BMP analogues. The entire CDR may be present in the small peptide BMP analogues of the invention, or an BMP-like portion of the CDR may comprise such small peptide BMP analogues. Two or more CDRs may be joined in "head-to-head", "head-to-tail", or "tail-to-tail" orientation to form dimer or multimer small peptide BMP analogues. Since each BMP-like monoclonal antibody may contain multiple CDRs, a single CDR may be present in the small peptide BMP analogue or different BMP-like CDRs from one or more BMP-like monoclonal antibodies may be present. Non-naturally occurring, synthetic, and D-amino acids may be substituted for specific amino acids of the BMP-like CDRs.

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The invention further encompasses peptides which specifically bind the truncated receptor and have BMP-like activity. These peptides may be, but are not exclusively, based on the sequence of the CDRs of BMP-like antibodies. The peptides may be "bridged" or "joined" to other peptides in multimeric structures of two peptides or more to elicit the BMP-like activity. The amino acids of the peptides may be, but are not exclusively, "unnatural" amino acids of, e.g., D-stereo-specificity or analogues of amino acids with other chemical groups attached to the peptide backbone. The peptides may also be cyclic in structure. A peptide as used herein is a molecule comprising of at least two amino acids and up to thirty amino acids.

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The invention further encompasses small organic molecules, which may include amino acid-like molecules, which exhibit specific binding to the truncated human BMP receptor and which possess BMP-like activity. "Small organic molecules" are defined in accordance with the present invention as non-protein carbon-containing molecules of molecular weight less than 3000 which have been first identified for use as BMP analogues using the truncated receptor of the invention. The small organic molecules of the invention are capable of being incorporated into oral pharmaceuticals for treatment of bone and/or cartilage disorders.

Pharmaceutical compositions containing the BMP-like monoclonal antibodies, small peptide BMP analogues, or small organic molecules of the present invention are useful in treating a variety of bone and/or cartilage disorders of diverse etiologies. Such pharmaceutical compositions may also contain pharmaceutically acceptable carriers,

diluents, fillers, salts, buffers, stabilizers, and/or other materials well known in the art. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier or other material will depend on the route of administration.

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Administration of the BMP-like monoclonal antibodies, small peptide BMP analogues or small organic molecules of the invention can be carried out in a variety of conventional ways, including via matrices and/or carriers such as are disclosed in U.S. Patent 5,171,579. For the BMP-like monoclonal antibodies, intravenous administration to the patient is preferred. Cutaneous or subcutaneous injection may also be employed for administration of the monoclonal antibody embodiment of the invention. Oral administration is preferred for administration of the small peptide and small organic molecule BMP analogue embodiments of the invention.

The amount of BMP-like monoclonal antibody, small peptide analogue or small organic molecule in the pharmaceutical composition of the present invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of BMP analogue with which to treat each individual patient. It is contemplated that the various pharmaceutical compositions of the present invention should contain about $0.1~\mu g$ to about $100~\mu g$ of BMP analogue molecule per kg body weight.

20 EXAMPLES

Example I. Identification of the murine KDA-B5 sequence.

Two peptide sequences were derived from the amino acid sequence of the activin receptor in the region described as the kinase domain of the molecule (Mathews et al., Cell, 65:973-982(1991)). These particular sequences were selected on the basis of a comparison between the amino acid sequence of the Daf-1 gene product and the activin receptor and are predicted to be conserved in other serine threonine kinase receptor molecules. The two peptide sequences selected were:

- 1. Asn-Glu-Tyr-Val-Ala-Val-Lys
- 2. His-Arg-Asp-Ile-Lys-Ser

The following oligonucleotide primer was designed on the basis of amino acid

sequence #1 and synthesized on an automated DNA synthesizer.

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Oligonucleotide primer A: GCGGATCCGARTAYGTNGCNGTNAAR

The first 8 nucleotides of this primer (underlined) comprise a recognition sequence for the restriction endonuclease BamHI in order to facilitate subsequent manipulations of amplified DNA products and are not derived from amino acid sequence #1.

The following oligonucleotide primers were designed on the basis of amino acid sequence #2 and synthesized on an automated DNA synthesizer.

Oligonucleotide primer B: GACTCTAGARCTYTTDATRTCYCTRTG

Oligonucleotide primer C: <u>GACTCTAGARCTYTTDATRTCNCGRTG</u>

Oligonucleotide primer D: GACTCTAGANGAYTTDATRTCYCTRTG

Oligonucleotide primer E: GACTCTAGANGAYTTDATRTCNCGRTG

The first 9 nucleotides of primers B through E (underlined) comprise a recognition sequence for the restriction endonuclease XbaI in order to facilitate subsequent manipulations of amplified DNA products and are not derived from amino acid sequence #2.

The standard nucleotide symbols in the above identified primers are as follows: A=adenosine, C=cytosine, G=guanine, T=thymine, R=adenosine or guanine, Y=cytosine or thymine and D=guanine, adenosine or thymine.

These oligonucleotides have been selected for their predicted ability to specifically amplify serine/threonine kinase domain encoding sequences similar to those found in the activin receptor sequence. Since activin and the BMP molecules are members of the large $TGF-\beta$ superfamily of growth and differentiation factors we predict that their corresponding receptors may also be related to each other in structure and primary amino acid sequence. The $TGF-\beta$ type II receptor sequence (Lin et al., Cell, 68:775-785 (1992)) indicates that like the activin receptor it is also a serine threonine kinase. On the basis of the above described relationships, we predicted that these degenerate oligonucleotides will specifically amplify sequences encoding fragments of other serine/threonine kinase receptor molecules including activin receptors, $TGF-\beta$ receptors and BMP receptors.

The BMP-2 responsive mouse cell line W-20-17 was selected as a source of mRNA which we would predict to contain molecules that are capable of encoding BMP receptors.

Total RNA was extracted from W-20-17 cells using established procedures known to those skilled in the art, and mRNA was subsequently selected for by oligo (dT) cellulose chromatography. 10 ng of the W-20-17 mRNA was utilized as a template to synthesize first strand cDNA in a reaction mixture (20 μ l total volume) containing 1 mM each deoxynucleotide triphosphate (dATP, dGTP, dCTP, dTTP), 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 5 mM MgCl₂, 1 U/ μ l RNase inhibitor, 2.5 U/ μ l reverse transcriptase, 2.5 μ M random hexmers and 20 ng of the W-20-17 mRNA described above. This reaction mixture was incubated for 10 minutes at room temperature, followed by 15 minutes at 42°C and then 5 minutes at 99°C. The completed first strand cDNA reaction was then placed at 4°C.

Oligonucleotide combinations consisting of oligonucleotide primer A paired with either oligonucleotide primer B, C, D or E were utilized as primers to allow the amplification of specific nucleotide sequences from the first strand W-20-17 cDNA template described above. The amplification reaction was performed by adjusting the first strand cDNA reaction described above (20 μ l) to a volume of 100 μ l in order to bring the components of the reaction buffer to the following final concentrations: 2 mM MgCl₂, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 2.5 U/100 μ l Taq DNA polymerase, 1 pM/ μ l oligonucleotide primer A and 1 pM/ μ l of either oligonucleotide primer B, C, D or E. The entire reaction mixture was then incubated at 95°C for two minutes and then subjected to thermal cycling in the following manner: 1 minute at 95°C, 1 minute at 40°C and 1 minute at 72°C for forty cycles; followed by a 7 minute incubation at 72°C, after which the completed reaction is held at 4°C.

The DNA which was specifically amplified by this reaction was ethanol precipitated, digested with the restriction endonucleases BamHI and XbaI and subjected to agarose gel electrophoresis. Regions of the gel in which DNA bands were evident were excised and the DNA contained within was eluted with a QIAEX Gel Extraction Kit (Qiagen catalog no: 20020) according to the instructions supplied by the manufacturer. The gel-extracted DNA fragments were subcloned into the plasmid vector pGEM-3 between the BamHI and XbaI sites of the polylinker. DNA sequence analysis of one of the resulting subclones named KDA-B5 indicated that the specifically amplified DNA

sequence insert encodes an amino acid sequence homologous to the corresponding region of the activin receptor and Daf-1 gene product kinase domains in the region between where the oligonucleotide primers A, B, C, D and E were designed (as described in the beginning of this section). The amino acid sequence encoded by this region of the specifically amplified KDA-B5 sequence is 41% and 47% identical to the corresponding regions of the activin receptor and Daf-1 kinase domains, respectively.

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The DNA sequence and derived amino acid sequence of the specifically amplified KDA-B5 DNA fragment are set forth in SEQ ID NO:11 and SEQ ID NO:12, respectively.

Nucleotides 1-24 of the sequence set forth in SEQ ID NO:11 comprise a portion of the oligonucleotide primer A and nucleotides 319-341 comprise a portion of the reverse compliment of oligonucleotide primer B utilized to perform the specific amplification reaction. Due to the function of oligonucleotides A and B in initiating the amplification reaction, they may not correspond exactly to the actual sequence encoding the mouse KDA-B5 protein and are therefore not translated in the above amino acid derivation.

Example II. Isolation of W101 and W120 clones from a W-20-17 cDNA library.

The 341 bp sequence of the KDA-B5 insert set forth in SEQ ID NO:11 was utilized as a probe to screen a W-20-17 cDNA library under reduced stringency conditions (4X SSC, 0.1% SDS at 60°C) in an attempt to isolate other mouse sequences related to KDA-B5 in the following manner.

1,000,000 recombinants of a W-20-17 (Thies et al., Endocrinology, 130:1318-1324 (1992)) cDNA library constructed in the vector λ ZAPII were plated at a density of 20,000 recombinant bacteriophage plaques per plate on 100 plates. Duplicate nitrocellulose replicas of the plates were made. A DNA fragment corresponding to the 341 bp sequence of the KDA-B5 insert set forth in SEQ ID NO:11 was ³²P-labelled by the random priming procedure of Feinberg et al., Anal. Biochem. 132:6-13 (1983), and hybridized to one set of filters in standard hybridization buffer (SHB: 5X SSC, 0.1% SDS, 5X Denhardt's, 100 μ g/ml salmon sperm DNA) at 60°C for 2 days. The other set of filters was hybridized to a DNA probe corresponding to nucleotides # 710 to 1044 of the published sequence of the activin receptor (Mathews et al., Cell, 65:973-982 (1991)) under the same conditions described for the first set. This region of the activin receptor kinase domain corresponds

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to the DNA sequence of the KDA-B5 insert. The filters were washed under reduced stringency conditions (4X SSC, 0.1% SDS at 60°C). 13 positively hybridizing recombinant bacteriophage plaques were selected and replated for secondaries. Duplicate nitrocellulose replicas of the recombinant plaques from these plates were made. Again. one set of filters was hybridized to the KDA-B5 probe and the other set to the activin receptor probe as described above in the primary screen (in SHB at 60°C for 2 days). Both sets of filters were washed under the reduced stringency conditions described above (4X SSC, 0.1% SDS at 60°C). Two recombinants which hybridized strongly to the KDA-B5 probe but not to the corresponding activin receptor probe were selected for further analysis. These two cDNA clones, designated W-101 and W-120, were plaque purified and their inserts were transferred to the plasmid Bluescript SK (+/-) according to the in vivo excision protocol described by the manufacturer (Stratagene). DNA sequence analysis of these recombinants indicated that they encode proteins homologous to the activin receptor and the Daf-1 gene product. The DNA sequence and derived amino acid sequence of a portion of pMT101 (ATCC 69379) is set forth in SEQ ID NO:7 and SEQ ID NO:8, respectively.

The nucleotide sequence of clone W-101 indicates that it encodes a partial polypeptide of 467 amino acids comprising the carboxy-terminal portion of the murine receptor molecule W-101. A random primed cDNA library was made from W-20-17 mRNA using a random hexmer pd(N)₆ (Pharmacia/LKB catalog # 27-2166-01) to prime synthesis of first strand cDNA from the W-20-17 mRNA template. The library was screened with a 30 base oligonucleotide corresponding to nucleotide # 245 to 274 of SEQ ID NO:7. The DNA sequence utilized to design this oligonucleotide probe was derived from the coding sequence of clone W-101. Hybridization was performed in SHB at 65°C and stringent wash conditions of 0.2X SSC, 0.1% SDS AT 65°C were employed to remove non-specifically bound probe. The DNA insert of one of these clones, WR9, was characterized by DNA sequence analysis and determined to contain additional 5' coding sequences of the murine W-101 receptor not present in the original cDNA clone described above. The 0.7 kb insert of the W9 clone however lacks sequences which encode the C-terminal region of the murine W-101 receptor protein. In order to construct a cDNA

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sequence which would encode the complete W-101 receptor protein, DNA sequences from both the original oligo (dT) and the random primed clone W9 have been joined at a common BstEII restriction endonuclease site. The sequence set forth in SEQ ID NO:7 contains an open reading frame of 1515 base pairs which encodes the complete 505 amino acid murine W-101 receptor protein. Nucleotides 1-660 of the sequence set forth in SEQ ID NO:7 are derived from the W9 cDNA clone while the remaining sequence (nucleotides 661-1648 are derived from the oligo (dT) primed cDNA clone W-101. The BstEII restriction endonuclease recognition sequence GGTNACC (located at position 660-666) facilitated the construction of this chimeric cDNA sequence. This construction was accomplished by digesting clone W-101 and clone W9 with the restriction endonuclease BstEII resulting in the linearization of each plasmid at the common BstEII site of their respective inserts. The two linearized plasmids were gel isolated, then ligated together at this site and digested with the restriction endonuclease SalI in order to separate the sequence set forth in SEQ ID NO:7 from the plasmid vector sequences (pBluescript). The DNA fragments resulting from this sequential ligation and SalI digest were electrophoresed on an agarose gel. A region containing a DNA fragment of approximately 2.3 kb. comprising 660 bp of the 5' end of the W9 cDNA and approximately 1.64 kb of the 3' end of the original W-101 cDNA, was excised from the gel and the DNA contained therein was eluted with a QIAEX Gel Extraction Kit (Qiagen catalog no: 20020) according to the instructions supplied by the manufacturer. The resulting DNA fragment which comprises the sequence set forth in SEQ ID NO:7 (encoding the complete murine W101 receptor protein) was subcloned into the mammalian cell expression vector pMT3 at the Sal I site of the polylinker region. This plasmid is designated pMT101.

The nucleotide sequence of a portion of the insert of cDNA clone W120 is set forth in SEQ ID NO:9. The presumed initiator methionine encoding sequence is preceded by 68 bp of 5' untranslated sequence and defines an open reading frame of 1509 bp which encodes the complete 503 amino acid murine receptor molecule W-120. The stop codon is followed by at least 203 bp of 3' untranslated sequence. The insert of clone W120 comprising the sequence set forth in SEQ ID NO:9 is excised from the pBluescript plasmid with the restriction endonuclease EcoRI and transferred to the mammalian cell expression

vector pMT3 at the EcoRI site of the polylinker region. This plasmid is designated pMT120E and has been deposited with the American Type Culture Collection (ATCC #69377).

Example III. Isolation of CFK1-10a, CFK1-23a and CFK1-43a from a CFK1 cDNA library.

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Another BMP-2 responsive cell line CFK1 [Bernier and Goltzman, J. Cell. Physiol., 152:317 (1992)] was selected as a source of mRNA which would be predicted to contain molecules capable of encoding BMP receptors and additional serine/threonine kinase encoding sequences related to the TGF- β receptor, the activin receptor, daf-1, W-101 and W-120.

1 x 10⁶ recombinants of a CFK1 cDNA library constructed in the vector λZAPII were plated at a density of 20,000 recombinant bacteriophage plaques per plate on 50 plates. Duplicate nitrocellulose replicas of the plates were made. A 645 base pair DNA fragment of the W-101 cDNA insert corresponding to nucleotides #828-#1472 of SEQ ID NO:7 was ³²P-labelled by the random priming procedure of Feinberg et al., Anal. Biochem., 132:6-13 (1983) and hybridized to both sets of filters in SHB at 60°C for two days. The filters were washed under reduced stringency conditions (4X SSC, 0.1% SDS at 60°C). Many duplicate hybridizing recombinants of various intensities (approximately 200) were noted. 27 bacteriophage plaques which were representative of the broad range of hybridization intensity were plaque purified and their inserts were transferred to the plasmid pBluescript SK (+/-) according to the in vivo excision protocol described by the manufacturer (Stratagene). DNA sequence analysis of several recombinants indicated that they encode proteins homologous to the activin receptor, *Daf-1* and other receptor proteins of the serine/threonine kinase receptor family.

The nucleotide sequence of clone CFK1-10a comprises an open reading frame of 1527 bp, encoding a CFK1-10a receptor protein of 509 amino acids. The encoded 509 amino acid CFK1-10a receptor protein is contemplated to be the primary translation product, as the coding sequence is preceded by 458 bp of 5' untranslated sequence with stop codons in all three reading frames. The DNA and derived amino acid sequence of the majority of the insert of CFK1-10a (ATCC # 69380) is set forth in SEQ ID NO:5.

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Based on the knowledge of other serine/threonine kinase receptor proteins.the encoded 509 amino acid CFK1-10a has the characteristic features of serine/threonine kinase receptors, particularly those capable of recognizing ligands of the TGF-\(\beta\)/BMP superfamily of growth and differentiation factors. This molecule encodes a full length receptor molecule with a characteristic hydrophobic leader sequence which targets the ligand binding domain of the protein to the extracellular space, a transmembrane region which anchors the complete receptor molecule in the cell membrane allowing the positioning of the serine threonine kinase domain within the intracellular space. The ligand binding domain of the CFK1-10a receptor molecule of the invention exhibits a pattern of cysteine conservation noted for other receptors capable of recognizing ligands of the TGF-β/BMP superfamily of growth and differentiation factors. The region of the CFK1-10 a receptor protein corresponding to the intracellular serine threonine kinase domain exhibits a significant degree of amino acid sequence identity to the corresponding domain of other receptors of this family as follows. TGF- β type II receptor (Genbank Accession No. M85079), 35%; activin type II receptor (Genbank Accession No. M65287), 40%; activin type IIB receptor (Genbank Accession No. M84120), 38%; and Daf-1 (Genbank Accession No. A35103), 39%. The 3.2 kb insert of the CFK1-10a cDNA clone is excised with the restriction endonuclease NotI and transferred to the mammalian cell expression vector pMV2 at the NotI site of the polylinker. This plasmid is designated CFK1-10a/Not-4.

The CFK1-10a receptor of the present invention is homologous to a human serine/threonine kinase receptor mRNA entered into Genbank, accession number L02911, for which no ligand was identified. This receptor protein is also homologous to the reported murine type I TGF- β receptor, Ebner et al., *Science*, 260:1344-1348 (1993)(Genbank Accession No. L15436).

The nucleotide sequence of clone CFK1-23a (ATCC # 69378) comprises an open reading frame of 1596 bp, encoding a CFK1-23a receptor protein of 532 amino acids. The encoded 532 amino acid CFK1-23a receptor protein is contemplated to be the primary translation product. The coding sequence is preceded by 60 bp of 5' untranslated sequence. The DNA and derived amino acid sequence of the majority of the insert of

CFK1-23a is set forth in SEO ID NO:1.

Based on the knowledge of other serine/threonine kinase receptor proteins, the encoded 532 amino acid CFK1-23a has the characteristic features of serine/threonine kinase receptors, particularly those capable of recognizing ligands of the TGF-β/BMP superfamily of growth and differentiation factors. The region of the CFK1-23a receptor protein corresponding to the intracellular serine threonine kinase domain exhibits a significant degree of amino acid sequence identity to the corresponding domain of other receptors of this family as follows: TGF-β type II receptor (Genbank Accession No. M85079), 35%; activin type II receptor (Genbank Accession No. M65287), 41%; activin type IIB receptor (Genbank Accession No. M84120), 39%; and Daf-1 (Genbank Accession No. A35103), 39%. The 2.8 kb insert of the CFK1-23a cDNA clone is excised with the restriction endonuclease EcoRI and transferred to the mammalian cell expression vector pMV2 at the EcoRI site of the polylinker. This plasmid is designated pMV23a.

The nucleotide sequence of clone CFK1-43a comprises an open reading frame of 1506 bp, encoding a CFK1-43a receptor protein of 502 amino acids. The encoded 502 amino acid CFK1-43a receptor protein is contemplated to be the primary translation product, as the coding sequence is preceded by 239 bp of 5' untranslated sequence with stop codons in all three reading frames. The DNA and derived amino acid sequence of the insert of CFK1-43a (ATCC # 69381) is set forth in SEO ID NO:3.

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Based on the knowledge of other serine/threonine kinase receptor proteins, the encoded 502 amino acid CFK1-43a has the characteristic features of serine/threonine kinase receptors, particularly those capable of recognizing ligands of the TGF- β /BMP superfamily of growth and differentiation factors. The region of the CFK1-43a receptor protein corresponding to the intracellular serine threonine kinase domain exhibits a significant degree of amino acid sequence identity to the corresponding domain of other receptors of this family as follows: TGF- β type II receptor (Genbank Accession No. M85079), 35%; activin type II receptor (Genbank Accession No. M65287), 41%; activin type IIB receptor (Genbank Accession No. M84120), 40%; and *Daf-1* (Genbank Accession No. A35103), 38%. The 2.1 kb insert of the CFK1-43a cDNA clone is excised with the restriction endonuclease EcoRI and transferred to the mammalian cell expression vector

pMV2 at the EcoRI site of the polylinker. This plasmid is designated pMV43a.

The CFK1-43a receptor of the invention is homologous to a chicken serine/threonine kinase receptor mRNA, entered into Genbank Accession No. D 13432, for which no ligand has been identified.

Example IV. Screening for Human BMP receptors.

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Mouse and/or rat BMP receptor genes are presumed to be significantly homologous. Therefore, the mouse coding sequences of W-101 or W-120 or portions thereof can be used to screen a human genomic or human cDNA library or as probes to identify a human cell line or tissue which synthesizes the analogous human BMP receptor proteins. In a similar manner, the rat coding sequences of CFK1-10a, CFK1-23a or CFK1-43a or portions thereof can be utilized to screen human libraries or as probes to identify a human cell line or tissue which synthesize the analogous human BMP receptor proteins. A human genomic library may be screened with such probes, and presumptive positively hybridizing recombinant clones isolated and DNA sequence obtained. Evidence that such recombinants encode portions of the corresponding human BMP receptor proteins relies on the murine or rat/human DNA, protein and gene structure homologies.

Once a recombinant bacteriophage or plasmid containing DNA encoding a portion of a human BMP receptor molecule is obtained, the human coding sequence can be used to identify a human cell line or tissue which synthesizes the corresponding human BMP receptor mRNA. Alternatively, the mouse or rat BMP receptor encoding sequence can be utilized as a probe to identify such a human cell line or tissue. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from a coding sequence from the mouse, rat or human BMP receptor. Alternatively, the mouse or rat BMP receptor coding sequence is used to design oligonucleotide primers which will specifically amplify a portion of the BMP receptor encoding sequence located in the region between the primers utilized to perform the specific amplification reaction. It is contemplated that mouse, rat and human BMP receptor coding sequences would be sufficiently homologous to allow one to specifically

amplify corresponding human BMP receptor encoding sequences from mRNA, cDNA or genomic DNA templates. Once a positive source has been identified by one of these above described methods, mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in a suitable vector (ie. λ gt10, λ ZAPII or other vectors known to those skilled in the art) by established techniques. It is also possible to perform the oligonucleotide primer-directed amplification reaction, described above, directly on a pre-established human cDNA or genomic library which has been cloned into a λ bacteriophage or plasmid vector. In such cases, a library which yields a specifically amplified DNA product encoding a portion of human BMP receptor protein could be screened directly, utilizing the fragment of amplified BMP receptor encoding DNA as a probe.

Example V. Expression of BMP receptors.

In order to produce mouse, rat, human or other mammalian BMP receptor proteins, the DNA encoding it is transferred into an appropriate expression vector (as described above for W-101, W-120, CFK1-10a, CFK1-23a and CFK1-43a) and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The DNA sequences encoding BMP receptor protein may be inserted into a vector suitable for a particular host cell, as described above. The preferred expression system for recombinant human BMP receptor proteins is contemplated to be stably transformed mammalian cells.

A. COS Cell Expression

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As one specific example of expressing a serine/threonine kinase receptor protein of the invention, the insert of CFK1-23a (containing the full length BMP receptor cDNA for CFK1-23a) is released from the vector arms by digestion with EcoRI and subcloned into the EcoRI site of the mammalian expression vector, pMV2, a derivative of pMT2, which has been deposited with ATCC wunder the accession number ATCC 67122, though other derivatives thereof, such as pMT3, may also be suitable. Plasmid DNA from this subclone is transfected into COS cells by the DEAE-dextran procedure [Sompayrac and Danna PNAS 78:7575-7578 (1981); Luthman and Magnusson, Nucl. Acids Res. 11: 1295-1308 (1983)] and the cells are cultured. Serum-free 24 hr. conditioned medium is

collected from the cells starting 40-70 hr. post-transfection.

B. CHO Cell Expression

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(1) Serine/threonine kinase Receptor Expression in CHO Cells

In order to achieve high levels of serine/threonine kinase receptor protein expression, each of 5the DNA sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7 or SEQ ID NO:9 are inserted into a eucaryotic expression vector, i.e., pMV2 or pMT3, stably introduced into CHO cells and amplified to high copy number by methotrexate selection of DHFR [Kaufman et al., EMBO L 6:189 (1987)]. The transformed cells are cultured and the expressed receptor proteins remain associated with the cell membrane of the transformed cell. Recombinant receptor proteins of the 10present invention can be dissociated from the transformed cell membrane and then are recovered and purified from other contaminants present.

A serine/threonine kinase receptor protein of the invention is expressed in CHO cells by releasing the insert of cloned CFK1-43a described above with EcoRI. The insert is subcloned into the EcoRI cloning site of the mammalian expression vector, pMV2 described above, though derivatives 15thereof, i.e., pMT3, may also be suitable.

Methods for producing heterologous protein from CHO cells are known in the art and are described above, at pages 16 through 19.

Example VI. Binding assays to determine affinity of cloned receptors for different TGF- β /BMP superfamily ligands.

A BMP receptor of the invention can be defined by a protein possessing the ability to bind a particular BMP at a greater binding affinity than TGF-β, activin, inhibin or other members of the TGF-β family of growth and differentiation factors. The BMP receptors of the present invention bind specifically to a particular BMP such that approximately a 100-fold excess of a competitive ligand such as TGF-β or activin will not significantly displace the BMP. Specific binding of BMPs to a particular 25BMP receptor of the invention can be demonstrated by transfecting an expression plasmid containing DNA sequences encoding the particular BMP receptor protein of interest into COS cells and allowing for the transient expression of the BMP receptor protein on the cell surface. Individual BMPs, heterodimeric BMPs or other proteins of the TGF-β superfamily are bound to BMP receptor expressing COS cells and the cells are analyzed for their ability to bind specifically to a BMP molecule 30or particular set of BMP proteins with greater affinity than to TGF-β or other members of the TGF-β

superfamily. Such binding assays may be performed in the following manner:

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COS cells that have been transfected with an expression vector containing the particular BMP receptor coding sequence of interest (e.g., pMT101, pMT120, CFK1-10a/Not-4, pMV23a or pMV43a) are plated on gelatinized 6 well plates and preincubated at 37°C for 60 minutes in binding buffer (128 mM NaCl, 5 mM KCl, 5 mM MgSO₄, 1.2 mM CaCl², 50 mM HEPES and 5 mg/ml BSA, pH 7.5). The preincubated COS cells are washed and incubated in binding buffer supplemented with 10 mM KCN and 2 mM NaF for 10 minutes prior to the addition of BMP-4 and/or [125 I]BMP-4. BMP-4 binding is allowed to equilibrate at 37°C for 60 minutes. Following binding, the cells are washed twice with ice-cold binding buffer and solubilized with 1% Triton X-100, 10% glycerol, 25 mM HEPES and 1 mg/ml BSA, pH 7.5, as described by Massague [Meth. Enzymol. 146: 174-195 (1987)]. Radioactivity is then determined in a gamma counter.

EXAMPLE VII. Production of truncated receptor proteins for production of truncated protein.

Truncated receptor proteins of the invention preferably comprise the ligand binding domain but not the transmembrane and serine/threonine kinase domains of the receptor proteins. Such truncated receptor proteins can be expressed in mammalian cells in a manner that the truncated receptor proteins will be secreted into the supernatant rather than be expressed on the surface of the host cell. DNA sequences encoding the ligand binding domain of each receptor protein of the invention can be isolated from DNA sequences encoding the transmembrane and serine/threonine kinase domains of each corresponding receptor protein of the invention and inserted into vectors which will allow for the production of truncated receptor proteins in mammalian cells. Alternatively, the DNA sequences encoding the truncated receptor proteins may be isolated and inserted into suitable vectors for expression in bacterial, insect, viral and yeast cells. Such vectors are known to those skilled in the art and are described elsewhere in this application.

In a preferred embodiment, DNA sequences comprising nucleotides # 61 through # 507 of SEQ ID NO:1 encoding amino acids # 1 through 149 of the CFK1-23a receptor protein of the invention (SEQ ID NO:2) can be specifically amplified and the resulting DNA sequence can be inserted into a standard mammalian cell expression vector (i.e.,

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pMV2 or pMT3). This specific amplification can be performed in the following manner: Oligonucleotide primers comprising the nucleotide sequence # 1 through # 20 of SEO ID NO:1 and a separate primer comprising the complimentary strand of nucleotide sequence # 488 through 507 of SEQ ID NO:1 are synthesized on an automated DNA synthesizer. Additionally these oligonucleotides could be designed to include recognition sequences of restriction endonucleases known by those skilled in the art (ie. BamHI, EcoRI, XbaI etc.) to be useful in the manipulation of amplified DNA products and the facilitation of their insertion into plasmid vectors. Furthermore the primer comprising nucleotides # 488 through 507 could also include a trinucleotide sequence corresponding to a translational stop codon (ie. TAA, TAG or TGA) in place of nucleotides # 583 through 585 of SEO ID NO:1. The oligonucleotide comprising nucleotides # 488 through 507 would be designed on the basis of the antisense (complementary) strand of this region of SEQ ID NO: 1. and in combination with an oligonucleotide comprising nucleotides # 1 through 20 of the sense (coding) strand of SEQ ID NO: 1 could be used to specifically amplify a DNA fragment comprising nucleotides #1 through # 507 of SEO ID NO: 1. This DNA fragment will encode a truncated receptor protein of the invention. The DNA fragment encoding the truncated receptor protein of the invention can be produced by specifically amplifying the sequence comprising nucleotides # 1 through # 507 of SEO ID NO: 1 through the use of a clone encoding the complete receptor of the invention, such as pMV23a, as a template. This specific DNA amplification reaction can be performed as follows: approximately 1 ng of template DNA, such as pMV23a is combined with 100 pM of an oligonucleotide comprising nucleotides #1 through 20 and 100 pM of an oligonucleotide comprising the complementary strand of nucleotides # 488 through 507 in a 100 µl reaction mixture consisting of 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl₂ 200 µM each deoxynucleotide triphosphate and 2.5 units of Taq DNA polymerase. The entire reaction is then incubated at 95° C for two minutes and then subjected tom thermal cycling in the following manner: 1 minute at 95° C, 1 minute at 40° C and 1 minute at 72° C for twenty-five to forty cycles; followed by a 7 minute incubation at 72° C, after which the completed reaction is held at 4° C.

The DNA fragment which is specifically amplified by this reaction is ethanol

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precipitated, digested with the appropriate restriction endonucleases in the cases where restriction endonuclease recognition sequences have been added to the oligonucleotides utilized to prime the synthesis of the amplified DNA fragment (described above), and subjected to agarose electrophoresis. A region of the gel in which a DNA band of the expected size is evident is excised and subcloned into a plasmid vector. Alternatively this specifically amplified DNA fragment encoding a truncated receptor protein of the invention could be subclone directly into a standard mammalian cell expression vector such as pMT3 or other such vectors known to those skilled in the art.

Similar manipulations could be performed as above, in order to isolate and express other truncated receptor proteins of the invention.

For example, DNA sequences comprising nucleotides #247 through 618 of SEQ ID NO:3 encoding amino acids # 1 through 124 of the CFK1-43a BMP receptor of the invention (SEO ID NO:4) can be specifically amplified to produce a DNA fragment comprising nucleotides # 247 through 618 of SEO ID NO:3 which will encode another truncated BMP receptor protein. These sequences will encode the ligand binding domain of the CFK1-43a BMP receptor but will not encode the transmembrane and serine/threonine kinase domains of the corresponding receptor protein. When inserted into an appropriate expression vector and transfected into the appropriate host cell, this construct will allow the production of a truncated BMP receptor protein which will be secreted into the medium rather than be expressed on the surface of the host cell. This specific amplification reaction can be performed in a similar manner to that described above with respect to the truncated CFK1-23a BMP receptor protein. In this case, oligonucleotides comprising the nucleotide sequence #247 through 266 of SEO ID NO:3 and a separate oligonucleotide primer comprising the complementary strand of nucleotide sequence # 599 through 618 of SEQ ID NO:3 are utilized to specifically amplify a DNA fragment comprising nucleotides # 247 through 618 of SEQ ID NO:3. oligonucleotides and a template DNA encoding the corresponding BMP receptor protein of the invention such as pMV43a, are substituted in the specific DNA amplification reaction mixture described earlier.

Additionally, other serine/threonine kinase receptors of the invention such as W-

101, W-120 or CFK1-10a can be produced in a truncated form. the truncated forms of these receptor molecules can be expressed in mammalian cells in a manner that the corresponding truncated proteins will be secreted into the culture media rather than be expressed on the surface of the host cell. The expression of these soluble receptor proteins can be accomplished through the amplification of DNA fragments encoding the ligand binding domain, but not the transmembrane and serine/threonine kinase domains, of each respective serine/threonine kinase receptor molecule as described above for the truncated BMP receptor proteins.

Example VIII. W-20-17 Bioassays

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A. <u>Description of W-20-17 cells</u>

Use of the W-20-17 bone marrow stromal cells as an indicator cell line is based upon the conversion of these cells to osteoblast-like cells after treatment with a BMP protein [Thies et al, Journal of Bone and Mineral Research, 5:305 (1990); and Thies et al, Endocrinology, 130:1318 (1992)]. Specifically, W-20-17 cells are a clonal bone marrow stromal cell line derived from adult mice by researchers in the laboratory of Dr. D. Nathan, Children's Hospital, Boston, MA. Treatment of W-20-17 cells with certain BMP proteins results in (1) increased alkaline phosphatase production, (2) induction of PTH stimulated cAMP, and (3) induction of osteocalcin synthesis by the cells. While (1) and (2) represent characteristics associated with the osteoblast phenotype, the ability to synthesize osteocalcin is a phenotypic property only displayed by mature osteoblasts. Furthermore, to date we have observed conversion of W-20-17 stromal cells to osteoblast-like cells only upon treatment with BMPs. In this manner, the in vitro activities displayed by BMP treated W-20-17 cells correlate with the in vivo bone forming activity known for BMPs.

Below two in vitro assays useful in comparison of BMP activities of formulations of BMPs with the activity of known BMPs are described.

B. W-20-17 Alkaline Phosphatase Assay Protocol

W-20-17 cells are plated into 96 well tissue culture plates at a density of 10,000 cells per well in 200 μ l of media (DME with 10% heat inactivated fetal calf serum, 2 mM glutamine and 100 Units/ml penicillin + 100 μ g/ml streptomycin. The cells are allowed

to attach overnight in a 95% air, 5% CO₂ incubator at 37°C.

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The 200 μ l of media is removed from each well with a multichannel pipettor and replaced with an equal volume of test sample delivered in DME with 10% heat inactivated fetal calf serum, 2 mM glutamine and 1% penicillin-streptomycin. Test substances are assayed in triplicate.

The test samples and standards are allowed a 24 hour incubation period with the W-20-17 indicator cells. After the 24 hours, plates are removed from the 37°C incubator and the test media are removed from the cells.

The W-20-17 cell layers are washed 3 times with 200 μ l per well of calcium/magnesium free phosphate buffered saline and these washes are discarded.

 $50 \mu l$ of glass distilled water is added to each well and the assay plates are then placed on a dry ice/ethanol bath for quick freezing. Once frozen, the assay plates are removed from the dry ice/ethanol bath and thawed at 37° C. This step is repeated 2 more times for a total of 3 freeze-thaw procedures. Once complete, the membrane bound alkaline phosphatase is available for measurement.

50 μ l of assay mix (50 mM glycine, 0.05% Triton X-100, 4 mM MgCl₂, 5 mM p-nitrophenol phosphate, pH = 10.3) is added to each assay well and the assay plates are then incubated for 30 minutes at 37°C in a shaking waterbath at 60 oscillations per minute.

At the end of the 30 minute incubation, the reaction is stopped by adding 100 μ l of 0.2 N NaOH to each well and placing the assay plates on ice.

The spectrophotometric absorbance for each well is read at a wavelength of 405 nanometers. These values are then compared to known standards to give an estimate of the alkaline phosphatase activity in each sample. For example, using known amounts of p-nitrophenol phosphate, absorbance values are generated. This is shown in Table 2.

25 Table 2

Absorbance values for known amounts of BMPs can be determined and converted to μ moles of p-nitrophenol phosphate cleaved per unit time as shown in Table 3.

30 Table 3

Values for Known Standards Nitrophenol Phosphate
Mean absorbance (405 nm)
0
0.261 +/024
0.521 +/031
0.797 +/063
1.074 +/061
1.305 +/083

All	caline Phosphatase Value Treating with E		lls
BMP-2 concentration ng/ml 4	Absorbance Read	ling um per hour	oles substrate
0	0.645	0.02	24
1.56	0.696	0.02	26
3.12	0.765	0.02	29
6.25	0.923	0.03	36
12.50	1.121	0.044	
25.0	1.457	0.03	58
50.0	1.662	0.06	57
100.0	1.977	0.080	

These values are then used to compare the activities of known amounts of new BMP formulations to known active BMP formulations.

C. Osteocalcin RIA Protocol

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W-20-17 cells are plated at 106 cells per well in 24 well multiwell tissue culture

dishes in 2 mls of DME containing 10% heat inactivated fetal calf serum, 2 mM glutamine. The cells are allowed to attach overnight in an atmosphere of 95% air 5% CO₂ at 37°C.

The next day the medium is changed to DME containing 10% fetal calf serum, 2 mM glutamine and the test substance in a total volume of 2 ml. Each test substance is administered to triplicate wells. The test substances are incubated with the W-20-17 cells for a total of 96 hours with replacement at 48 hours by the same test medias.

At the end of 96 hours, 50 μ l of the test media is removed from each well and assayed for osteocalcin production using a radioimmunoassay for mouse osteocalcin. The details of the assay are described in the kit manufactured by Biomedical Technologies Inc., 378 Page Street, Stoughton, MA 02072. Reagents for the assay are found as product numbers BT-431 (mouse osteocalcin standard), BT-432 (Goat anti-mouse Osteocalcin), BT-431R (iodinated mouse osteocalcin), BT-415 (normal goat serum) and BT-414 (donkey anti goat IgG). The RIA for osteocalcin synthesized by W-20-17 cells in response to BMP treatment is carried out as described in the protocol provided by the manufacturer.

Example IX. Rat Ectopic Study

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Twenty four Long-Evans male rats are divided into 6 test groups. Each receives a subcutaneous implant, 200 uL in size, with either a 0 or 20 ug/100 uL dose of a particular BMP/matrix sample, for example BMP/PLGA porous particles/blood clot, as disclosed in U.S. Patent 5,171,579, the disclosure of which is hereby incorporated by reference. After 14 days, the rats are sacrificed and each animal is evaluated for bone formation.

Example X. Polyclonal Antibodies against the Truncated BMP Receptor

Three rabbits are injected with purified truncated BMP receptor protein. Polyclonal antibodies from sera of these rabbits are purified by chromatography on a Protein A column. The antibodies are tested for the ability to bind BMP receptors.

Example XI. Monoclonal Antibodies Against the Truncated Human BMP Receptor

Three sets of three mice are immunized with purified truncated BMP receptor protein. The spleens of the mice are used to generate multiple hybridoma cell lines. The

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conditioned media from non-clonal mouse hybridoma lines are tested for ability to bind to BMP receptors. Clonal lines can be developed from these by sequential serial and limiting dilution cloning. As non-clonal lines become clonal those lines which are positive for activity in the non-clonal stage will retain the ability to produce an antibody which binds BMP receptor. Conversely, lines which are negative will stay negative throughout the cloning process. Monoclonal antibodies may be purified from ascites derived from both positive and negative hybridoma clones.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: GENETICS INSTITUTE, INC.
- (ii) TITLE OF INVENTION: RECEPTOR PROTEINS
- (iii) NUMBER OF SEQUENCES: 19
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Genetics Institute Inc. Legal Affairs
 - (B) STREET: 87 CambridgePark Drive
 - (C) CITY: Cambridge
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 - (E) COUNTRY: USA
 - (F) ZIP: 02140
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: TBD
 - (B) FILING DATE: HEREWITH
 - (C) CLASSIFICATION
- (vii PRIOR APPLICATION DATA
 - (A) APPLICATION NUMBER: US 08/123,934
 - (B) FILING DATE: 17-SEP-1993
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: LAZAR, Steven R
 - (B) REGISTRATION NUMBER: 32,618
 - (C) REFERENCE/DOCKET NUMBER: 5203-PCT
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (617) 498-8260
 - (B) TELEFAX: (617) 876-5851
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1813 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: CFK1-23a
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 61..1656

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

	/ * *	•	_	CD D.												
CTA	GTGG.	ATC	CCCC	GGGC'	TG C	AGGA	ATTC!	T GC	GGCC	GCCA	GGA	CACG	TGC	GAAT	TGGACi	A 60
				TAC Tyr 5												108
				GTT Val											GGT Gly	156
			Lys	TCA Ser												204
				GAG Glu												252
				GAT Asp												300
				ATT Ile 85												348
				ATG Met												396
				CAG Gln												444
				TAT Tyr												492
				GGC Gly												540
				GTC Val 165												588
				AAG Lys												636
				GAA Glu												684
				TCA Ser												732
				ACT Thr												780
				TAT Tyr 245												828

AA Ly:	A GTO	G GC: l Ala	r GTC a Val 260	Lys	A GTI	A TTI L Phe	TTI Phe	T ACC	r Thi	r GAA	A GAI	A GCT 1 Ala	AGC Sei 270	Tr	F TTT Phe	876
AG:	A GAI g Glu	A ACI Thi 275	Glu	ATC Ile	TAC Tyr	CAG Gln	ACC Thr 280	· Val	TT!	A ATO	CG:	CAT His 285	Glu	AA: Asi	T ATA	924
CT: Lei	GG1 G13 290	, bh∈	TATA	GCT Ala	GCA Ala	GAC Asp 295	Ile	AAA Lys	GGC Gly	ACC Thr	300 300	ser Ser	TGG	ACT Thi	CAG Gln	972
Let 305	Tyr	Leu	Ile	Thr	Asp 310	Tyr	His	Glu	Asn	315	Ser	Leu	Tyr	Asp	TTC Phe 320	1020
CTC Lev	AAA Lys	TGT Cys	GCC Ala	ACC Thr 325	Leu	GAC Asp	ACC Thr	AGA Arg	GCC Ala 330	Leu	CTC Leu	AAG Lys	TTA Leu	GCT Ala 335	TAT	1068
TCI Ser	GCT Ala	GCC Ala	TGT Cys 340	Gly	CTG Leu	TGC Cys	CAC	CTC Leu 345	His	ACA Thr	GAA Glu	ATT Ile	TAT Tyr 350	GGC Gly	ACG Thr	1116
CAA Gln	GGC Gly	Lys 355	CCT Pro	GCA Ala	ATT Ile	GCT Ala	CAT His 360	CGA Arg	GAC Asp	CTG Leu	AAG Lys	AGC Ser 365	AAA Lys	AAC Asn	ATC Ile	1164
CTT Leu	ATT Ile 370	Lys	AAA Lys	AAT Asn	GGT Gly	AGT Ser 375	TGC Cys	TGT Cys	ATT Ile	GCT Ala	GAC Asp 380	CTG Leu	GGC Gly	CTA Leu	GCT Ala	1212
GTT Val 385	Lys	TTC Phe	AAC Asn	AGT Ser	GAC Asp 390	ACA Thr	TAA Asn	GAA Glu	GTT Val	GAC Asp 395	ATA Ile	CCC Pro	TTG Leu	AAC Asn	ACC Thr 400	1260
AGG Arg	GTG Val	GGC Gly	ACC Thr	AGG Arg 405	CGG Arg	TAC Tyr	ATG Met	GCT Ala	CCA Pro 410	GAA Glu	GTG Val	CTG Leu	GAC Asp	GAG Glu 415	AGC Ser	1308
Leu	Ser	Lys	AAC Asn 420	His	Phe	Gln	Pro	Tyr 425	Ile	Met	Ala	Asp	Ile 430	Tyr	Ser	1356
TTT Phe	GGT Gly	TTG Leu 435	ATC Ile	ATT Ile	TGG Trp	GAG Glu	ATG Met 440	GCC Ala	CGT Arg	CGC Arg	TGT Cys	ATT Ile 445	ACA Thr	GGA Gly	GGA Gly	1404
ATC Ile	GTG Val 450	GAG Glu	GAA Glu	TAT Tyr	CAA Gln	TTA Leu 455	CCA Pro	TAT Tyr	TAC Tyr	AAC Asn	ATG Met 460	GTG Val	CCT Pro	AGT Ser	GAC Asp	1452
CCA Pro 465	TCT Ser	TAT Tyr	GAA Glu	GAC Asp	ATG Met 470	CGT Arg	GAG Glu	GTC Val	GTG Val	TGT Cys 475	GTG Val	AAA Lys	CGC Arg	TTG Leu	CGG Arg 480	1500
CCA Pro	ATC Ile	GTC Val	TCT Ser	AAC Asn 485	CGC Arg	TGG Trp	AAC Asn	AGT Ser	GAT Asp 490	GAA Glu	TGT Cys	CTT Leu	CGA Arg	GCC Ala 495	GTT Val	1548
TTG Leu	AAG Lys	CTG Leu	ATG Met 500	TCA Ser	GAA Glu	TGC Cys	Trp	GCC Ala 505	CAT His	AAT Asn	CCA Pro	Ala	TCC Ser 510	AGA Arg	CTC Leu	1596
ACA Thr	Ala	TTG Leu 515	AGA . Arg	ATC Ile	AAG Lys	Lys	ACG Thr 520	CTC Leu	GCA Ala	AAG Lys	Met	GTT Val 525	GAA Glu	TCC Ser	CAG Gln	1644

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GAT GTA AAG ATT TGACAAACAG TTTTGAGAAA GAATTTAGAC TGCAAGAAAT Asp Val Lys Ile

TCACCCGAGG AAGGGTGGAG TTAGCATGGA CTAGGATGTC GGCTTGGTTT CCAGACTCTC 1756 TCCTCTACCA TCTTCACAGG CTGCTAACAG TAAACCTTTC AGGACTCTGC AGAATGC 1813

1696

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 532 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Thr Gln Leu Tyr Thr Tyr Ile Arg Leu Leu Gly Ala Cys Leu Phe

Ile Ile Ser His Val Gln Gly Gln Asn Leu Asp Ser Met Leu His Gly

Thr Gly Met Lys Ser Asp Val Asp Gln Lys Lys Pro Glu Asn Gly Val

Thr Leu Ala Pro Glu Asp Thr Leu Pro Phe Leu Lys Cys Tyr Cys Ser

Gly His Cys Pro Asp Asp Ala Ile Asn Asn Thr Cys Ile Thr Asn Gly

His Cys Phe Ala Ile Ile Glu Glu Asp Asp Gln Gly Glu Thr Thr Leu

Thr Ser Gly Cys Met Lys Tyr Glu Gly Ser Asp Phe Gln Cys Lys Asp

Ser Pro Lys Ala Gln Leu Arg Arg Thr Ile Glu Cys Cys Arg Thr Asn

Leu Cys Asn Gln Tyr Leu Gln Pro Thr Leu Pro Pro Val Val Ile Gly

Pro Phe Phe Asp Gly Ser Val Arg Trp Leu Ala Val Leu Ile Ser Met 155

Ala Val Cys Ile Val Ala Met Ile Val Phe Ser Ser Cys Phe Cys Tyr

Lys His Tyr Cys Lys Ser Ile Ser Ser Arg Gly Arg Tyr Asn Arg Asp

Leu Glu Gln Asp Glu Ala Phe Ile Pro Val Gly Glu Ser Leu Lys Asp 200

Leu Ile Asp Gln Ser Gln Ser Ser Gly Ser Gly Leu Pro Leu

Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Arg Gln Val 235

Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu 250

Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser Trp Phe 260 265 270

Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile 275 280 285

Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp Thr Gln 290 295 300

Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr Asp Phe 305 310 315 320

Leu Lys Cys Ala Thr Leu Asp Thr Arg Ala Leu Leu Lys Leu Ala Tyr 325 330 335

Ser Ala Ala Cys Gly Leu Cys His Leu His Thr Glu Ile Tyr Gly Thr 340 345 350

Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile 355 360 365

Leu Ile Lys Lys Asn Gly Ser Cys Cys Ile Ala Asp Leu Gly Leu Ala 370 380

Val Lys Phe Asn Ser Asp Thr Asn Glu Val Asp Ile Pro Leu Asn Thr 385 390 395 400

Arg Val Gly Thr Arg Arg Tyr Met Ala Pro Glu Val Leu Asp Glu Ser 405 410 415

Leu Ser Lys Asn His Phe Gln Pro Tyr Ile Met Ala Asp Ile Tyr Ser 420 425 430

Phe Gly Leu Ile Ile Trp Glu Met Ala Arg Arg Cys Ile Thr Gly Gly 435

Ile Val Glu Glu Tyr Gln Leu Pro Tyr Tyr Asn Met Val Pro Ser Asp 450 455 460

Pro Ser Tyr Glu Asp Met Arg Glu Val Val Cys Val Lys Arg Leu Arg 465 470 475 480

Pro Ile Val Ser Asn Arg Trp Asn Ser Asp Glu Cys Leu Arg Ala Val 485 490 495

Leu Lys Leu Met Ser Glu Cys Trp Ala His Asn Pro Ala Ser Arg Leu 500 505 510

Thr Ala Leu Arg Ile Lys Lys Thr Leu Ala Lys Met Val Glu Ser Gln 515 520 525

Asp Val Lys Ile 530

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2076 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: CFK1-43a

PCT/US94/10080 WO 95/07982

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 247..1752

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

	(XI) SE	ÕOEN	ע פט	ESCR	1P11	ON:	SEQ	ID N							
GCG	GCCG	CGC	CGGC	GTGG	TG C	TCGG	agtg	C GG	GCGC	CGAG	GAC	CCGG	GAC	CAGG	GCCCC	60
GCG	GCGG	GTT	GGAG	TTCA	AG G	TACT	CGTT	A CG	TGTG	ACGA	GGA	AGTG	AAG	CCCA	TTCCAT	120
GCC	TTGC	TGA	GAAA	GGTT	CA A	ACTT	CGGC	T GA	ATCA	CAAC	CAT	TTGG	CGC	TGAG	CTATGA	180
CAA	GAGA	GCA .	AACA	AAAA	GT T	AAAG	GAGC.	A AC	TCGG	CCAT	AAG	TGAC	AGA	GAAG	TTCGTT	240
GAT										TTA :						288
	Glu									GCT Ala 25						336
										GAC Asp						384
				Gly						ATA Ile						432
										GGA Gly			Gly			480
		Сув								CAG Gln		Arg				528
										GAT Asp 105						576
										GGA Gly						624
										TTA Leu						672
										CAA Gln						720
										TAC Tyr						768
										AGC Ser 185						816
										GCT Ala						864

	CAA Gln							AAG Lys		912
	GGA Gly 225							GAA Glu		960
 	TGG Trp					-				1008
 	AAC Asn	 	 		 			 		1056
	ACT Thr								;	1104
	GAC Asp								;	1152
	GCC Ala 305								:	1200
	AGC Ser								:	1248
	AAC Asn								1	1296
	CTG Leu								1	1344
	AAC Asn								נ	1392
 	GAG Glu 385	 	 	 	 		 	 	1	440
	TAC Tyr								3	.488
	GGA Gly								3	.536
	AGT Ser								1	.584
	TTA Leu								1	.632
	CAA Gln 465								1	.680

				ACA CTT GCC Thr Leu Ala 490		1728
	r Gln Asp 1		TGACGTCAGG	TACTTGTGGA C	AGAGCAAGG	1782
AATTACACAG	AAGCATCCTI	AGCCCAAGC	C TTGAACGTTG	ATCTACTGCC	CAGTGAGTTC	1842
AGACTTTCCT	CTAAGAGAGC	AAGCTGGAC	A GACACAGAGG	AACCCAGAAA	CACGGCTTCA	1902
CCATGGCTTT	CTGAGGAGGG	GAAACCATT	r gggtaacttg	TTCAAGATAT (GATGCATGTT	1962
GCTTTCTAAG	AAAGCCCTGT	ATTTTGGGA	TACCATTTTT	TTTAAAGAAG	AAAGATACTT	2022
TAATTTTTAC	CAAAATAAAA	CAAATATTA	DAAAAAAAG	CGGCCGCAGA	ATTC	2076

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 502 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Leu Leu Arg Ser Ser Gly Lys Leu Asn Val Gly Thr Lys Lys Glu
1 5 10 15

Asp Gly Glu Ser Thr Ala Pro Thr Ala Arg Pro Lys Val Leu Arg Cys 20 25 30

Lys Cys His His Cys Pro Glu Asp Ser Val Asn Asn Ile Cys Ser 35 40 45

Thr Asp Gly Tyr Cys Phe Thr Met Ile Glu Glu Asp Asp Ser Gly Thr
50 55 60

Pro Val Val Thr Ser Gly Cys Leu Gly Leu Glu Gly Ser Asp Phe Gln 65 70 75 80

Cys Arg Asp Thr Pro Ile Pro His Gln Arg Arg Ser Ile Glu Cys Cys 85 90 95

Thr Glu Arg Asn Glu Cys Asn Lys Asp Leu His Pro Thr Leu Pro Pro 100 105 110

Leu Lys Asp Arg Asp Phe Val Asp Gly Pro Ile His His Lys Ala Leu 115 120 125

Leu Ile Ser Val Thr Val Cys Ser Leu Leu Leu Val Leu Ile Ile Leu 130 135 140

Phe Cys Tyr Phe Arg Tyr Lys Arg Gln Glu Ala Arg Pro Arg Tyr Ser 145 150 155 160

Ile Gly Leu Glu Gln Asp Glu Thr Tyr Ile Pro Pro Gly Glu Ser Leu 165 170 175

Arg Asp Leu Ile Glu Gln Ser Gln Ser Ser Gly Ser Gly Leu 180 185 190

Pro Leu Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Lys Gln Ile Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser 235 Trp Phe Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp Thr Gln Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr Asp Tyr Leu Lys Ser Thr Thr Leu Asp Ala Lys Ser Met Leu Lys Leu Ala Tyr Ser Ser Val Ser Gly Leu Cys His Leu His Thr Glu Ile Phe Ser Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Thr Cys Cys Ile Ala Asp Leu Gly Leu Ala Val Lys Phe Ile Ser Asp Thr Asn Glu Val Asp Ile Pro Pro Asn Thr Arg Val Gly Thr Lys Arg Tyr Met Pro Pro Glu Val Leu Asp Glu Ser Leu Asn Arg Thr His Phe Gln Ser Tyr Ile Met Ala Asp Met Tyr Ser Phe Gly Leu Ile Leu Trp Glu Ile Ala Arg Arg Cys Val Ser Gly Gly Ile Val Glu Glu Tyr Gln Leu Pro Tyr His Asp Leu Val Pro Ser Asp Pro Ser Tyr Glu Asp Met Arg Glu Ile Val Cys Met Lys Lys Leu Arg Pro Ser Phe Pro Asn Arg Trp Ser Ser Asp Glu Cys Leu Arg Gln Met Gly Lys Leu Met Thr Glu Cys Trp Ala His Asn Pro Ala Ser Arg Leu Thr Ala Leu Arg Val Lys Lys Thr Leu Ala Lys Met Ser Glu 485 490

Ser Gln Asp Ile Lys Leu 500

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3238 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(vii) IMMEDIATE SOURCE: (B) CLONE: CFK1-10a

(ix) FEATURE:

(A) NAME/KEY: CDS (B) LOCATION: 474..2000

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

	•	•	_								-					
GA	ATTC:	rgcg	GCC	CGA	GC 1	rgcai	TAAG	T G	GAT	ATGC	AC	CCGT	GATT	CTG	ACAGC	CG 60
TG	ACTG	CGTG	GAG	CTG	CTC C	GGAA	CTCT	C CA	CAG	GGAG	CAI	AAGG	AGCT	GCC	CTCTG1	NG 120
TC	rccc	GCC	CTT	CAGCO	SAG A	GTCI	GGAA	A GA	GAAC	CGAG	GTO	CTA	CTGC	AGT	ggatg <i>i</i>	AG 180
TAC	BAGAJ	GAG	TCT	CATO	CCA G	TGCI	GGTG	A GC	TTGI	CTGG	CT	ATAGO	GGAG	CCT	CTGG	G 240
GA	ACTI	ACA	GCTI	CAG	AG A	CTCC	TGGA	G AG	CCTC	TCCC	TCC	CACAC	CTCT	ccc	DAGTTT	C 300
AGI	CAGI	GCC	TCTC	TGCI	GG A	GAAC	CTGT	G CT	GGGI	GTGC	ccc	AGAC	CTG	GCTI	TGACI	G 360
TAC	CCTG	TCA	GGCI	CTCC	CT G	GACC	TCAC	G GA	ACAG	CATT	GCC	AGC	CACA	CGGC	CTTCCA	A 420
CAA	ATCA	CCT	CTTT	TCAT	GC T	GTTT	GGCA	C AG	ATCG	AATC	TAC	AGGI	TAT	ACA	ATG Met 1	476
GTC Val	GAT Asp	GGA Gly	GCA Ala 5	ATG Met	ATC Ile	CTT Leu	TCT Ser	GTG Val 10	CTA Leu	ATG Met	ATG Met	ATG Met	GCT Ala 15	Leu	CCT Pro	524
TCC Ser	CCG Pro	AGT Ser 20	Met	GAA Glu	GAT	GAG Glu	GAG Glu 25	CCC Pro	AAG Lys	GTC Val	AAC Asn	CCG Pro	Lys	CTT	TAC	572
ATG Met	TGT Cys 35	Val	TGT Cys	GAG Glu	GGC Gly	CTC Leu 40	TCC Ser	TGC Cys	GGG Gly	AAC Asn	GAG Glu 45	GAC Asp	CAC His	TGT Cys	GAG Glu	620
GGC Gly 50	Gln	CAG Gln	TGT	TTT Phe	TCC Ser 55	TCC Ser	CTG Leu	AGC Ser	GTC Val	AAT Asn 00	yab	GGC	TTC Phe	CGC Arg	GTC Val 65	668
TAC Tyr	CAG Gln	AAG Lys	GGC Gly	TGC Cys 70	TTT Phe	CAG Gln	GTC Val	TAT Tyr	GAG Glu 75	CAG Gln	GGG Gly	AAG Lys	ATG Met	ACG Thr 80	TGT Cys	716
AAG Lys	ACC Thr	CCG Pro	CCG Pro 85	TCG Ser	CCT Pro	GGC Gly	CAG Gln	GCT Ala 90	GTG Val	GAG Glu	TGC Cys	TGC Cys	CAA Gln 95	GGG Gly	GAC Asp	764
TGG Trp	TGC Cys	Asn	Arg	Asn	Val	Thr	GCC Ala 105	Arg	Leu	Pro	Thr	Lvs	Glv	AAA Lys	TCC Ser	812
TTC Phe	CCT Pro 115	GGA Gly	TCG Ser	CAG Gln	AAC Asn	TTC Phe 120	CAC His	CTG Leu	GAA Glu	GTT Val	GGC Gly 125	CTT Leu	ATC Ile	ATC Ile	CTC Leu	860
TCC Ser 130	GTG Val	GTG Val	TTT Phe	GCG Ala	GTA Val 135	TGC Cys	CTT Leu	TTC Phe	GCT Ala	TGC Cys 140	ATC Ile	CTT Leu	GGC Gly	GTT Val	GCT Ala 145	908

CT(Let	AGC Arç	AAG J Lye	F TTI	AAA Lye 150	Arç	G CGC J Arg	TAA : naA :	CAA Glr	GAG Glu 155	Arç	CTC Lev	AA S	C CC	C AGI O Arg	A GAC 3 Asp	956
GT(Va)	GAG Glu	TAC Tyr	GGT Gly 165	Thr	ATC	GAA Glu	GGG Gly	CTC Leu 170	lle	ACC Thr	ACC Thr	AAC Asr	GTC Val 179	L Gly	A GAT	1004
AGC Ser	ACI Thr	CTA Leu 180	ı Ala	GAA Glu	TTA Leu	CTA Leu	GAT Asp 185	CAC	TCA Ser	TGI Cys	ACA Thr	TCA Ser 190	. G17	A AGI	GGC	1052
TCC Ser	GGT Gly 195	Leu	CCT Pro	TTT Phe	CTG Leu	GTA Val 200	CAG Gln	AGA Arg	ACT Thr	GTG Val	GCT Ala 205	Arg	CAG Glr	ATA lle	ACC Thr	1100
CTG Leu 210	Leu	GAG Glu	TGT Cys	GTC Val	GGG Gly 215	Lys	GGC	CGG Arg	TAT	GGA Gly 220	Glu	GTG Val	TGG	AGG Arg	GGC Gly 225	1148
AGC Ser	TGG Trp	CAA Gln	GGC Gly	GAA Glu 230	Asn	GTT Val	GCT Ala	GTG Val	AAG Lys 235	ATC	TTC Phe	TCC	TCC Ser	CGT Arg 240	Asp	1196
GAG Glu	AAG Lys	TCG Ser	TGG Trp 245	TTC Phe	AGG Arg	GAG Glu	ACA Thr	GAA Glu 250	TTG Leu	TAC Tyr	AAC Asn	ACG Thr	GTG Val 255	Met	CTG Leu	1244
AGG Arg	CAT His	GAG Glu 260	Asn	ATC Ile	TTA Leu	GGT	TTC Phe 265	ATT Ile	GCT Ala	TCA Ser	GAC Asp	ATG Met 270	Thr	TCT Ser	AGA Arg	1292
CAC His	TCC Ser 275	AGT Ser	ACC Thr	CAG Gln	CTG Leu	TGG Trp 280	CTC Leu	ATT Ile	ACA Thr	CAT His	TAC Tyr 285	CAC His	GAA Glu	ATG Met	GGA Gly	1340
Ser 290	Leu	Tyr	Asp	Tyr	Leu 295	CAG Gln	Leu	Thr	Thr	Leu 300	Asp	Thr	Val	Ser	Сув 305	1388
Leu	Arg	Ile	Val	10 310	Ser	ATA Ile	Ala	Ser	Gly 315	Leu	Ala	His	Leu	His 320	Ile	1436
Glu	Ile	Phe	Gly 325	Thr	Gln	GGG	Lys	Ser 330	Ala	Ile	Ala	His	Arg 335	Asp	Leu	1484
AAG Lys	AGC Ser	AAA Lys 340	AAC Asn	ATC Ile	CTC Leu	GTG Val	AAG Lys 345	AAG Lys	AAC Asn	GGA Gly	CAG Gln	TGC Cys 350	TGC Cys	ATA Ile	GCA Ala	1532
GAT Asp	TTG Leu 355	GGC Gly	CTG Leu	GCA Ala	GTC Val	ATG Met 360	CAT His	TCC Ser	CAG Gln	AGC Ser	ACG Thr 365	AAT Asn	CAG Gln	CTT Leu	GAT Asp	1580
GTG Val 370	GGA Gly	AAC Asn	AAC Asn	CCC Pro	CGT Arg 375	GTG Val	GGG Gly	ACC Thr	AAG Lys	CGC Arg 380	TAC Tyr	ATG Met	GCC Ala	CCT Pro	GAA Glu 385	1628
GTG Val	CTT Leu	GAT Asp	GAA Glu	ACC Thr 390	ATC Ile	CAA Gln	GTG Val	qaA	TGC Cys 395	TTT Phe	GAT Asp	TCT Ser	TAT Tyr	AAG Lys 400	AGG Arg	1676
GTC Val	GAT Asp	Ile	TGG Trp 405	GCC Ala	TTT Phe	GGC Gly	Leu '	GTT Val 410	CTG Leu	TGG Trp	GAA Glu	GTG Val	GCC Ala 415	AGG Arg	AGG Arg	1724

ATG GTG : Met Val	AGC Ser 420	AAT Asn	GGT Gly	ATA Ile	GTG Val	GAA Glu 425	GAT Asp	TAC Tyr	AAG Lys	CCA Pro	CCA Pro 430	TTC Phe	TAT Tyr	GAT Asp	1	.772
GTT GTT (Val Val Val 435	CCC Pro	TAA Asn	GAC Asp	CCA Pro	AGT Ser 440	TTT Phe	GAA Glu	GAT Asp	ATG Met	AGG Arg 445	AAA Lys	GTT Val	GTC Val	TGT Cys	1	.820
GTG GAT (Val Asp (450	CAA Gln	CAG Gln	AGG Arg	CCA Pro 455	AAC Asn	ATA Ile	CCT Pro	AAC Asn	AGA Arg 460	TGG Trp	TTC Phe	TCA Ser	GAC Asp	CCG Pro 465	1	868
ACA TTA I															1	916
CCA TCC	Ala														1	964
ATT GAT											TGAC	CATTO	GTC		2	010
ACCGGTGT	CA A	GAAG	GAGA	G TC	CAATO	CTGI	CAT	TGTC	CAG	CTGG	GACC	CTA I	ATGC	regect	2	070
GACTGGTT	GT C	AGAA	CAGA	A TO	CATO	TGTC	ccc	CTCI	ccc	CCCF	ACTO	cc c	GAAGI	GGCTG	2	130
CTTTGACA	AA A	GCAG	ATGI	C TO	TTCC	CAGO	CAT	GTTC	CGG	GGG	GACA	ACC 1	AAAA	CACCC	2	190
TAACCTCG	CT C	AGAA	ACTO	T GA	CTC	AGCA	CTI	GATG	AAC	TGTI	CACA	cc o	CAA	GACTA	2	250
ACGGTGGG	CA G	GTAI	GTTI	G CF	AGGG	GGAG	GGA	AGTG	GAG	GAGO	CACAC	AG 1	AGATO	CTGCA	2	310
GGAGATCT	GG G	CATT	AGGA	C AG	TGGC	TCTI	TGC	GTAI	CTT	CCAC	GGG1	CT (CTAC	ACTCG	2	370
CCCCACGG	GA A	ACTO	AAGG	A GO	CGGI	GAAI	TCG	LAAT	CAG	CAAT	OTTA:	GC 1	rgcgc	CTACT	2	430
CTTCTCTG:	TT G	CACI	AGGA	A TI	CTCI	GCAI	TCC	TTAC	TTG	CACI	GTCG	TC (TTA	TCTTA	2	490
AAGACCCG	AC T	TGCC	AAA	C AI	TGGC	TGCC	TAC	TTCA	CTG	GCCI	GTCI	CT (GACA	ATAGG	2	550
AATTCAAT	CT G	GCGA	AACA	AA AA	ATGI	DTAA'	TTG	GACI	TTG	CTGC	TTTA:	TA C	CACAC	GTGCC	2	610
GATGTTTA	CA A	CGAT	GCAA	A CA	TTAG	GAAI	TGI	TTAG	ACA	CAAC	TTTG	CA A	\ATTA	TTTAT	20	670
TACTGGTG	CA C	TTAG	CAGI	T TI	TGTI	TTTI	TTI	GTTI	TTT	TGTI	TTTT	TT 1	TGTI	TTGTT	2	730
TTGTTTTT	AT A	TATA	AAAC	T GC	CTCG	TGCG	TAT	'GTTA	AAG	CTTA	TTTT	TA 1	GTGG	TCTTA	2	790
TGATTTTA!	TT A	CCGA	AATG	T TI	TTAA	CACC	CGA	TTCT	GAA	ATGG	ATGI	TT I	CTTI	TATTA	28	B50
TCAGTTAA	AT T	CACA	TTTI	A AA	TGCI	TCAC	TTI	TTTT	TTA	TGTG	TGTA	GA C	TGTA	ACTTT	29	910
CTTTTCAG	TT A	GTAT	ACAG	A AC	GTAT	TTAG	CCA	TTAC	CCA	TGCA	ACAC	CA C	CCAA	TATAT	29	970
TACTGATT	TA G	AAGC	AAAG	A TI	TCAG	TAGA	ATT	TTAG	TCC	CAAA	CGCT	GT G	GGGG	GGAAA	30	030
TGCATCTT	CT T	CGGA	ACTA	T CC	ATTA	CATG	CAT	TTAA	ACT	CTGC	CAGA	AA A	AAAA	ATAAC	3(090
TATTTTGT	rr T	AATC	TACT	T TI	TGTA	TTTA	GTA	GTTA	TTT	GTAT	TAAA	TA A	ATAA	ACTGT	3:	150
TTTCAAGTO	CA A	AAAA	AAAA	A AA	AAAA	AAAA	AAA	AAAA	AAA	AAAA	AAAA	AA A	AAAA	TAAAA	32	210
AAAAAAA	AA AA	AAGC	GGCC	G CA	GAAT	TC									32	238

PCT/US94/10080 WO 95/07982

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 509 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Val Asp Gly Ala Met Ile Leu Ser Val Leu Met Met Ala Leu Pro Ser Pro Ser Met Glu Asp Glu Glu Pro Lys Val Asn Pro Lys Leu Tyr Met Cys Val Cys Glu Gly Leu Ser Cys Gly Asn Glu Asp His Cys Glu Gly Gln Gln Cys Phe Ser Ser Leu Ser Val Asn Asp Gly Phe Arg Val Tyr Gln Lys Gly Cys Phe Gln Val Tyr Glu Gln Gly Lys Met Thr Cys Lys Thr Pro Pro Ser Pro Gly Gln Ala Val Glu Cys Cys Gln Gly Asp Trp Cys Asn Arg Asn Val Thr Ala Arg Leu Pro Thr Lys Gly Lys Ser Phe Pro Gly Ser Gln Asn Phe His Leu Glu Val Gly Leu Ile Ile Leu Ser Val Val Phe Ala Val Cys Leu Phe Ala Cys Ile Leu Gly Val 135 Ala Leu Arg Lys Phe Lys Arg Arg Asn Gln Glu Arg Leu Asn Pro Arg Asp Val Glu Tyr Gly Thr Ile Glu Gly Leu Ile Thr Thr Asn Val Gly Asp Ser Thr Leu Ala Glu Leu Leu Asp His Ser Cys Thr Ser Gly Ser Gly Ser Gly Leu Pro Phe Leu Val Gln Arg Thr Val Ala Arg Gln Ile Thr Leu Leu Glu Cys Val Gly Lys Gly Arg Tyr Gly Glu Val Trp Arg Gly Ser Trp Gln Gly Glu Asn Val Ala Val Lys Ile Phe Ser Ser Arg Asp Glu Lys Ser Trp Phe Arg Glu Thr Glu Leu Tyr Asn Thr Val Met 250 Leu Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ser Asp Met Thr Ser 260 Arg His Ser Ser Thr Gln Leu Trp Leu Ile Thr His Tyr His Glu Met Gly Ser Leu Tyr Asp Tyr Leu Gln Leu Thr Thr Leu Asp Thr Val Ser

Сув 305	Leu	Arg	Ile	Val	Leu 310	Ser	Ile	Ala	Ser	Gly 315	Leu	Ala	His	Leu	His 320
Ile	Glu	Ile	Phe	Gly 325	Thr	Gln	Gly	Lys	Ser 330	Ala	Ile	Ala	His	Arg 335	Asp
Leu	Lys	Ser	Lys 340	Asn	Ile	Leu	Val	Lув 345	Lys	Asn	Gly	Gln	Сув 350	Сув	Ile
Ala	Asp	Leu 355	Gly	Leu	Ala	Val	Met 360	His	Ser	Gln	Ser	Thr 365	Asn	Gln	Leu
двр	Val 370	Gly	Asn	Asn	Pro	Arg 375	Val	Gly	Thr	Lys	Arg 380	Tyr	Met	Ala	Pro
Glu 385	Val	Leu	Asp	Glu	Thr 390	Ile	Gln	Val	Asp	Сув 395	Phe	Asp	Ser	Tyr	Lys 400
Arg	Val	Asp	Ile	Trp 405	Ala	Phe	Gly	Leu	Val 410	Leu	Trp	Glu	Val	Ala 415	
Arg	Met	Val	Ser 420	Asn	Gly	Ile	Val	Glu 425	Asp	Tyr	Lys	Pro	Pro 430	Phe	Tyr
Asp	Val	Val 435	Pro	Asn	Asp	Pro	Ser 440	Phe	Glu	Asp	Met	Arg 445	Lys	Val	Val
Сув	Val 450	Asp	Gln	Gln	Arg	Pro 455	Asn	Ile	Pro	Asn	Arg 460	Trp	Phe	Ser	Asp
Pro 465	Thr	Leu	Thr	Ser	Leu . 470	Ala	Lys	Leu	Met	Lys 475	Glu	Сув	Trp	Tyr	Gln 480
Asn	Pro	Ser	Ala	Arg 485	Leu	Thr	Ala	Leu	Arg 490	Ile	Lys	Lys	Thr	Leu 495	Thr
Lys	Ile	Asp	Asn 500	Ser	Leu	Asp	Lys	Leu 505	Lys	Thr	Asp	Сув			
(2)	INFO	RMAI	CION	FOR	SEQ	ID N	10:7:								
	(i)				IARAC				_						
		(E	3) TY	PE:	nucl	.eic	acid	l	8						
					EDNE			116							
	(ii)	MOI	.ECUI	E TY	PE:	DNA	(ger	omic	:)						

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 80..1594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

																	•
CCC Pro	CTT Leu	GTT Val	GTC Val 15	CTC Leu	CTG Leu	CTC Leu	GCC Ala	GGC Gly 20	AGC Ser	GGC Gly	GGG Gly	TCC Ser	GGG Gly 25	CCC Pro	CGG Arg		160
GGG Gly	ATC Ile	CAG Gln 30	GCT Ala	CTG Leu	CTG Leu	TGT Cys	GCG Ala 35	TGC Cys	ACC Thr	AGC Ser	TGC Cys	CTA Leu 40	CAG Gln	ACC Thr	AAC Asn		208
					GAT Asp												256
					CAT His 65												304
					CCC Pro												352
					TAT Tyr												400
					CTC Leu												448
					GGC Gly												496
					GTC Val 145												544
					AGG Arg												592
					AAG Lys												640
ACG Thr					TCA Ser												688
					TTA Leu												736
					CGC Arg 225												784
TTC Phe	TCT Ser	TCT Ser	CGT Arg	GAA Glu 240	GAA Glu	CGG Arg	TCT Ser	TGG Trp	TTC Phe 245	CGT Arg	GAA Glu	GCA Ala	GAG Glu	ATC Ile 250	TAC Tyr		832
					CGC Arg											,	880
GAC Asp	AAT Asn	AAA Lys 270	GAT Asp	AAT Aen	GGC Gly	ACC Thr	TGG Trp 275	ACC Thr	CAG Gln	CTG Leu	TGG Trp	CTT Leu 280	GTC Val	TCT Ser	GAC Asp	1	928

TAT Tyr	CAC His 285	Glu	CAT His	Gly	TCA Ser	Leu 290	TTT	GAT Asp	TAT	CTG Leu	AAC Asn 295	CGC Arg	TAC	ACC Thr	GTG Val	976
	Ile					Lys					Ala				TTG Leu 315	1024
GCA Ala	CAC His	CTG Leu	CAT His	ATG Met 320	GAG Glu	ATT Ile	GTG Val	GGC Gly	ACT Thr 325	CAA Gln	GGG	AAG Lys	CCG Pro	GGA Gly 330	ATT Ile	1072
GCT Ala	CAT His	CGA Arg	GAC Asp 335	TTG Leu	AAG Lys	TCA Ser	AAG Lys	AAC Asn 340	ATC Ile	CTG Leu	GTG Val	AAA Lys	AAA Lys 345	AAT Asn	GGC Gly	1120
ATG Met	TGT Cys	GCC Ala 350	ATT	GCA Ala	GAC Asp	CTG Leu	GGC Gly 355	CTG Leu	GCT Ala	GTC Val	CGT Arg	CAT His 360	GAT Asp	GCG Ala	GTC Val	1168
ACT Thr	GAC Asp 365	ACC Thr	ATA Ile	GAC Asp	ATT Ile	GCT Ala 370	CCA Pro	AAT Asn	CAG Gln	AGG Arg	GTG Val 375	GGG Gly	ACC Thr	AAA Lys	CGA Arg	1216
TAC Tyr 380	ATG Met	GCT Ala	CCT Pro	GAA Glu	GTC Val 385	CTT Leu	GAC Asp	GAG Glu	ACA Thr	ATC Ile 390	AAC Asn	ATG Met	AAG Lys	CAC His	TTT Phe 395	1264
Asp Asp	TCC Ser	TTC Phe	AAA Lys	TGT Cys 400	GCC Ala	GAC Asp	ATC Ile	TAT Tyr	GCC Ala 405	CTC Leu	GGG Gly	CTT Leu	GTC Val	TAC Tyr 410	TGG Trp	1312
GAG Glu	ATT Ile	GCA Ala	CGA Arg 415	AGA Arg	TGC Cys	AAT Asn	TCT Ser	GGA Gly 420	GGA Gly	GTC Val	CAT His	GAA Glu	GAC Asp 425	TAT Tyr	CAA Gln	1360
CTG Leu	CCG Pro	TAT Tyr 430	TAC Tyr	GAC Asp	TTA Leu	GTG Val	CCC Pro 435	TCC Ser	GAC Asp	CCT Pro	TCC Ser	ATT Ile 440	GAG Glu	GAG Glu	ATG Met	1408
CGA Arg	AAG Lys 445	GTT Val	GTA Val	TGT Cys	GAC Asp	CAG Gln 450	AAG Lys	CTA Leu	CGG Arg	CCC Pro	AAT Asn 455	GTC Val	CCC Pro	AAC Asn	TGG Trp	1456
TGG Trp 460	CAG Gln	AGT Ser	TAT Tyr	GAG Glu	GCC Ala 465	TTG Leu	CGA Arg	GTG Val	ATG Met	GGA Gly 470	AAG Lys	ATG Met	ATG Met	CGG Arg	GAG Glu 475	1504
TGC Cys	TGG Trp	TAC Tyr	GCC Ala	AAT Asn 480	GGT Gly	GCT Ala	GCC Ala	CGT Arg	CTG Leu 485	ACA Thr	GCT Ala	CTG Leu	CGC Arg	ATC Ile 490	AAG Lys	1552
AAG Lys	ACT Thr	CTG Leu	TCC Ser 495	CAG Gln	CTA Leu	AGC Ser	Val	CAG Gln 500	GAA Glu	GAT Asp	GTG Val	Lys	ATT Ile 505			1594
TAAG	CTGT	TA A	GATG	CCTA	C AC	AAAG	AACC	TGG	GCAG	TGA	GGAT	GACT	GC A	GG		1647

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 505 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ala Glu Ser Ala Gly Ala Ser Ser Phe Phe Pro Leu Val Val Leu Leu Leu Ala Gly Ser Gly Gly Ser Gly Pro Arg Gly Ile Gln Ala Leu Leu Cys Ala Cys Thr Ser Cys Leu Gln Thr Asn Tyr Thr Cys Glu Thr Asp Gly Ala Cys Met Val Ser Ile Phe Asn Leu Asp Gly Val Glu His His Val Arg Thr Cys Ile Pro Lys Val Glu Leu Val Pro Ala Gly Lys Pro Phe Tyr Cys Leu Ser Ser Glu Asp Leu Arg Asn Thr His Cys Cys Tyr Ile Asp Phe Cys Asn Lys Ile Asp Leu Arg Val Pro Ser Gly His 105 Leu Lys Glu Pro Ala His Pro Ser Met Trp Gly Pro Val Glu Leu Val Gly Ile Ile Ala Gly Pro Val Phe Leu Leu Phe Leu Ile Ile Ile Ile 135 Val Phe Leu Val Ile Asn Tyr His Gln Arg Val Tyr His Asn Arg Gln Arg Leu Asp Met Glu Asp Pro Ser Cys Glu Met Cys Leu Ser Lys Asp 165 Lys Thr Leu Gln Asp Leu Val Tyr Asp Leu Ser Thr Ser Gly Ser Gly Ser Gly Leu Pro Leu Phe Val Gln Arg Thr Val Ala Arg Thr Ile Val Leu Gln Glu Ile Ile Gly Lys Gly Arg Phe Gly Glu Val Trp Arg Gly Arg Trp Arg Gly Gly Asp Val Ala Val Lys Ile Phe Ser Ser Arg Glu Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile Tyr Gln Thr Val Met Leu Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Asn Lys Asp Asn Gly Thr Trp Thr Gln Leu Trp Leu Val Ser Asp Tyr His Glu His Gly 280 Ser Leu Phe Asp Tyr Leu Asn Arg Tyr Thr Val Thr Ile Glu Gly Met Ile Lys Leu Ala Leu Ser Ala Ala Ser Gly Leu Ala His Leu His Met Glu Ile Val Gly Thr Gln Gly Lys Pro Gly Ile Ala His Arg Asp Leu 330 Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Met Cys Ala Ile Ala

Asp	Leu	Gly 355	Leu	Ala	Val	Arg	His 360	Asp	Ala	Val	Thr	Авр 365	Thr	Ile	Asp	
Ile	Ala 370	Pro	Asn	Gln	Arg	Val 375	Gly	Thr	Lys	Arg	Tyr 380	Met	Ala	Pro	Glu	
Val 385	Leu	Asp	Glu	Thr	Ile 390	Asn	Met	Lув	His	Phe 395	Asp	Ser	Phe	Lys	Сув 400	
Ala	Asp	Ile	Tyr	Ala 405	Leu	Gly	Leu	Val	Tyr 410	Trp	Glu	Ile	Ala	Arg 415	Arg	
Сув	Asn	Ser	Gly 420	Gly	Val	His	Glu	Asp 425	Tyr	Gln	Leu	Pro	Tyr 430	Tyr	Asp	
Leu	Val	Pro 435	Ser	Asp	Pro	Ser	Ile 440	Glu	Glu	Met	Arg	Lys 445	Val	Val	Сув	
Asp	Gln 450	Lys	Leu	Arg	Pro	Asn 455	Val	Pro	Asn	Trp	Trp 460	Gln	Ser	Tyr	Glu	
Ala 465	Leu	Arg	Val	Met	Gly 470	Lys	Met	Met	Arg	Glu 475	Сув	Trp	Tyr	Ala	Asn 480	
Gly	Ala	Ala	Arg	Leu 485	Thr	Ala	Leu	Arg	Ile 490	Lys	Lys	Thr	Leu	Ser 495	Gln	
Leu	Ser	Val	Gln 500	Glu	Asp	Val	Lys	Ile 505								
(2)	INFO	RMAI	NOI	FOR	SEQ	ID N	10:9:	:								
	/ 5 1	SEC	QUENC	יות כיו	IDRAC	ומארי	בידר									
	(-)	(Ž) LE	ENGTH	1: 17	794 E	ase	pair	8							
		•	3) TY 3) SI													
) TC					,								
	(ii)	MOI	ECUI	E TY	PE:	DNA	(gen	omic	:)							
((vii)		ŒDIA 3) CI		_											
	(ix)		TURE													
			A) NA B) LO				1591									
		\-	,		. • • • • • • • • • • • • • • • • • • •			-								
	•		UENC					_								
GAAI	TCGC	GG C	CGCG	GGCG	A GG	CTTC	CTGA	GGA	GAAG	CTG	CGGC	CGGG	GC C	GGGC	CGGGC	60
CACA	AACA	GT G	GCGG	CGGG	A CC		Glu				Ala				CGT Arg 10	112
CCG Pro	CAG Gln	CTC Leu	CTC Leu	ATC Ile 15	GTG Val	TTG Leu	GTG Val	GCG Ala	GCG Ala 20	GCG Ala	ACG Thr	CTG Leu	CTC Leu	CCG Pro 25	GGG Gly	160
											ACA Thr					208

			Thr					Phe							ACA Thr	256
		Val				AGT Ser 65										304
	Arg					GTA Val										352
						TGC									GAA Glu	400
						TTT Phe										448
						GTC Val										496
						GTC Val 145										544
CAC His 155	CAC His	CGT Arg	GTG Val	CCA Pro	AAT Asn 160	GAA Glu	GAG Glu	GAT Asp	CCA Pro	TCA Ser 165	CTA Leu	GAT Asp	CGC Arg	CCT Pro	TTC Phe 170	592
						TTA Leu										640
						TTA Leu										688
AGG Arg	ACC Thr	ATT Ile 205	GTG Val	TTA Leu	CAA Gln	GAA Glu	AGC Ser 210	ATT Ile	GGC Gly	AAA Lys	GGT Gly	CGG Arg 215	TTT Phe	GGA Gly	GAA Glu	736
						CGG Arg 225										784
						TCA Ser										832
						GAA Glu										880
						TGG Trp										928
						TTC Phe										976
						CTT Leu 305										1024

	Leu					Val						CCA Pro				1072
												AAA Lys				1120
												GAT Asp				1168
GAT Asp	ACA Thr	ATT Ile 365	GAT Asp	ATT Ile	GCT Ala	CCA Pro	AAC Asn 370	CAC His	AGA Arg	GTA Val	GGC Gly	ACT Thr 375	AAA Lys	AGG Arg	TAC Tyr	1216
ATG Met	GCC Ala 380	CCT Pro	GAA Glu	GTT Val	CTA Leu	GAT Asp 385	GAT Asp	TCC Ser	ATA Ile	AAT Asn	ATG Met 390	AAA Lys	CAT His	TTT Phe	GAA Glu	1264
TCC Ser 395	TTC Phe	AAA Lys	CGC Arg	GCT Ala	GAC Asp 400	ATC Ile	TAT Tyr	GCA Ala	ATG Met	GGC Gly 405	TTA Leu	GTG Val	TTC Phe	TGG Trp	GAA Glu 410	1312
ATT Ile	GCT Ala	CGA Arg	CGC Arg	TGT Cys 415	TCT Ser	ATT Ile	GGT Gly	GGA Gly	ATC Ile 420	CAT His	GAA Glu	GAC Asp	TAT Tyr	CAG Gln 425	TTG Leu	1360
CCT Pro	TAT Tyr	TAT Tyr	GAT Asp 430	CTT Leu	GTA Val	CCT Pro	TCT Ser	GAT Asp 435	CCA Pro	TCG Ser	GTT Val	GAA Glu	GAA Glu 440	ATG Met	AGA Arg	1408
AAA Lys	GTA Val	GTT Val 445	TGC Cyb	GAA Glu	CAG Gln	AAG Lys	TTA Leu 450	AGG Arg	CCA Pro	AAT Asn	ATT Ile	CCA Pro 455	AAC Asn	AGA Arg	TGG Trp	1456
CAG Gln	AGC Ser 460	TGT Cys	GAG Glu	GCC Ala	TTG Leu	AGA Arg 465	GTG Val	ATG Met	GCT Ala	AAA Lys	ATT Ile 470	ATG Met	AGA Arg	GAA Glu	TGC Cys	1504
TGG Trp 475	TAT Tyr	GCC Ala	AAT Asn	GGA Gly	GCA Ala 480	GCA Ala	AGG Arg	CTG Leu	Thr	GCT Ala 485	TTG Leu	CGA Arg	ATT Ile	Lys	AAA Lys 490	1552
ACA Thr	TTG Leu	TCA Ser	Gln	CTC Leu 495	AGC Ser	CAA Gln	CAG Gln	Glu	GGC Gly 500	ATC Ile	AAA Lys	ATG Met	TAAC	TGAA	AC	1601
ACCG	TGGG	AA C	TCTG	CTCT	C TT	CATA	TCTG	CTC	CTGG	GTG	TTTA	GGAG	GC T	GGTT	GTTCT	1661
ACCT	CACT	GA G	AGAA	CAGA	G GG	CTCT	GCTT	CCT	CTTG	CAG	CAGT	GGAA'	TA T	GGTC	AACTG	1721
AAAG	CTTC	CC A	GGGT'	TTCT	C TG	GGCC	CAGA	GGC	AGCC	GTG	GGGT	CCTT	CT G	TGCA	CTATG	1781
GATA	ACTT	CT T	CC													1704

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 503 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Glu Ala Ala Ala Ala Pro Arg Arg Pro Gln Leu Leu Ile Val Leu Val Ala Ala Ala Thr Leu Leu Pro Gly Ala Lys Ala Leu Gln Cys Phe Cys His Leu Cys Thr Lys Asp Asn Phe Thr Cys Glu Thr Asp Gly Leu Cys Phe Val Ser Val Thr Glu Thr Thr Asp Lys Val Ile His Asn Ser Met Cys Ile Ala Glu Ile Asp Leu Ile Pro Arg Asp Arg Pro Phe Val Cys Ala Pro Ser Ser Lys Thr Gly Ala Val Thr Thr Thr Tyr Cys Cys Asn Gln Asp His Cys Asn Lys Ile Glu Leu Pro Thr Thr Gly Pro Phe Ser Glu Lys Gln Ser Ala Gly Leu Gly Pro Val Glu Leu Ala Ala Val Ile Ala Gly Pro Val Cys Phe Val Cys Ile Ala Leu Met Leu Met Val Tyr Ile Cys His Asn Arg Thr Val Ile His His Arg Val Pro Asn Glu Glu Asp Pro Ser Leu Asp Arg Pro Phe Ile Ser Glu Gly Thr Thr Leu Lys Asp Leu Ile Tyr Asp Met Thr Thr Ser Gly Ser Gly Leu Pro Leu Leu Val Gln Arg Thr Ile Ala Arg Thr Ile Val Leu Gln Glu Ser Ile Gly Lys Gly Arg Phe Gly Glu Val Trp Arg Gly Lys Trp Arg Gly Glu Glu Val Ala Val Lys Ile Phe Ser Ser Arg Glu Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile Tyr Gln Thr Val Met Leu Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Asn Lys Asp Asn Gly Thr Trp Thr Gln Leu Trp Leu Val Ser Asp Tyr His Glu His Gly Ser Leu Phe Asp Tyr Leu Asn Arg Tyr Thr Val Thr Val Glu Gly Met Ile Lys Leu Ala Leu Ser Thr Ala Ser Gly Leu Ala His Leu His Met Glu Ile Val Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser 330 Lys Asn Ile Leu Val Lys Lys Asn Gly Thr Cys Cys Ile Ala Asp Leu

Gly	Leu	Ala 355		Arg	His	Asp	Ser 360	Ala	Thr	Asp	Thr	Ile 365	Asp	Ile	Ala	
Pro	Asn 370		Arg	Val	Gly	Thr 375	Lys	Arg	Tyr	Met	Ala 380	Pro	Glu	Val	Leu	
А вр 385	Asp	Ser	Ile	Asn	Met 390	Lys	His	Phe	Glu	Ser 395	Phe	Lys	Arg	Ala	Asp 400	
Ile	Tyr	Ala	Met	Gly 405	Leu	Val	Phe	Trp	Glu 410	Ile	Ala	Arg	Arg	Cys 415	Ser	
Ile	Gly	Gly	Ile 420	His	Glu	Asp	Tyr	Gln 425	Leu	Pro	Tyr	Tyr	Asp 430	Leu	Val	
Pro	Ser	Asp 435	Pro	Ser	Val	Glu	Glu 440	Met	Arg	Lys	Val	Val 445	Cys	Glu	Gln	
Lys	Leu 450	Arg	Pro	Asn	Ile	Pro 455	Asn	Arg	Trp	Gln	Ser 460	Сув	Glu	Ala	Leu	
Arg 465	Val	Met	Ala	Lys	Ile 470	Met	Arg	Glu	Сув	Trp 475	Tyr	Ala	naA	Gly	Ala 480	
Ala	Arg	Leu	Thr	Ala 485	Leu	Arg	Ile	Lys	Lys 490	Thr	Leu	Ser	Gln	Leu 495	Ser	
Gln	Gln	Glu	Gly 500	Ile	Lys	Met										
(2)	INFO	RMAI	NOI	FOR	SEQ	ID N	0:11	. :								
	(i)	() (E) LE 3) TY 3) SI	NGTH PE: RAND	: 34 nucl	TERI 1 ba eic SS: line	se p acid sing	airs !	1							
	(ii)	MOI	ECUL	E TY	PE:	DNA	(gen	omic)							
(vii)		EDIA) CL													
	(ix)	(A	TURE () NA () LO	ME/K		CDS 25	318									
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:11:						
GGAT	CCGA	AT A	.CGTG	GCGG	T TA	AA A' I					GG G rg A					51
TGG Trp 10	TTC Phe	CGT Arg	GAG (Glu)	GCG Ala	GAA . Glu 15	ATT ! Ile !	TAT (CAG :	ACG (GTG : Val : 20	ATG (Met)	CTG :	AGA (CAC (GAG Glu 25	99

GAC Asp	TAT Tyr	TTG Leu 60	AAC Asn	AGA Arg	AAC Asn	ATA Ile	GTG Val 65	ACT Thr	GTG Val	GCT Ala	GGA Gly	ATG Met 70	GTC Val	AAG Lys	CTG Leu	243
GCG Ala	CTT Leu 75	TCC Ser	ATA Ile	GCG Ala	AGT Ser	GGT Gly 80	CTG Leu	GCT Ala	CAC His	CTG Leu	CAC His 85	ATG Met	GAG Glu	ATC Ile	GTG Val	291
			GGT Gly						CACC	GTG1	ATA 1	rcaa <i>i</i>	AAGT	CT		338
AGA				1												341

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 98 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ile Phe Ser Ser Arg Asp Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile 1 5 10 15

Tyr Gln Thr Val Met Leu Arg His Glu Asn Ile Leu Gly Phe Ile Ala 20 25 30

Ala Asp Asn Lys Asp Asn Gly Thr Trp Thr Gln Leu Trp Leu Val Ser 35 40 45

Glu Tyr His Glu Gln Gly Ser Leu Tyr Asp Tyr Leu Asn Arg Asn Ile 50 55 60

Val Thr Val Ala Gly Met Val Lys Leu Ala Leu Ser Ile Ala Ser Gly 65 70 75 80

Leu Ala His Leu His Met Glu Ile Val Gly Thr Gln Gly Lys Leu Ala 85 90 95

Ile Ala

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: PRIMER A
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCGGATCCGA RTAYGTNGCN GTNAAR

(2) INFORMATION FOR SEQ ID NO:14:

26

(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (A) LIBRARY: PRIMER B	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GACTGTAG	AR CTYTTDATRT CYCTRTG	27
(2) INFO	RMATION FOR SEQ ID NO:15:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (B) CLONE: PRIMER C	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:15:	
GACTCTAGA	AR CTYTTDATRT CNCGRTG	27
(2) INFOR	RMATION FOR SEQ ID NO:16:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(vii)	IMMEDIATE SOURCE: (B) CLONE: PRIMER D	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:16:	
GACTCTAGN	G AYTTDATRTC YCTRTG	26
(2) INFOR	MATION FOR SEQ ID NO:17:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) 1	MOLECULE TYPE: DNA (genomic)	
(2244)	THAT TATE COUDER.	

(B) CLONE: PRIMER E

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GACTCTAGAN GAYTTDATRT CNCGRTG

27

- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: PEPTIDE SEQUENCE OF KDA-B5 USED TO DESIGN PRIMER A
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Asn Glu Tyr Val Ala Val Lys 5

- (2) INFORMATION FOR SEQ ID NO:19:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 6 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: PEPTIDE SEQUENCE OF KDA-B5 USED TO DESIGN PRIMERS B THRU E
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

His Arg Asp Ile Lys Ser

What is claimed is:

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1. An isolated DNA molecule comprising a DNA sequence encoding a BMP receptor protein.

- 2. The DNA molecule of claim 1, wherein said DNA sequence is selected from the group consisting of:
 - (a) nucleotides 61 to 1656 of SEQ ID NO: 1;
 - (b) nucleotides 247 to 1752 of SEQ ID NO: 3; and
- (c) sequences which hybridize to (a) or (b) under stringent hybridization conditions and encode a BMP receptor protein with the ability to bind to BMP in a binding assay.
- 3. An isolated DNA molecule comprising a DNA sequence encoding a serine/threonine kinase receptor protein, wherein the DNA sequence is selected from the group consisting of:
 - (a) nucleotides 474 to 2000 of SEQ ID NO: 5;
 - (b) nucleotides 80 to 1594 of SEQ ID NO: 7;
 - (c) nucleotides 83 to 1591 of SEQ ID NO: 9;
- (d) sequences which hybridize to any of (c) to (e) under stringent hybridization conditions and encode a serine/threonine receptor protein.
- 4. The DNA molecule of claim 1, wherein said DNA sequence is selected from the group consisting of:
 - (a) nucleotides encoding for amino acids 24 to 532 of SEQ ID NO: 2; and
 - (b) nucleotides encoding for amino acids 8 to 502 of SEQ ID NO: 4; and
- (c) sequences which hybridize to any of (a) or (b) under stringent hybridization conditions and encode a BMP receptor protein with the ability to bind to BMP in a binding assay.
- 5. An isolated DNA molecule comprising a DNA sequence encoding a serine/kinase receptor protein, wherein the DNA sequence is selected from the group consisting of:
 - (a) nucleotides encoding for amino acids 18 to 509 of SEQ ID NO: 6; and
- 30 (b) nucleotides encoding for amino acids 24 to 505 of SEQ ID NO: 8; and
 - (c) nucleotides encoding for amino acids 30 to 503 of SEQ ID NO:10; and

(d) sequences which hybridize to any of (a) to (c) under stringent hybridization conditions and encode a serine/threonine kinase receptor protein.

- 6. A host cell transformed with the DNA molecule of claim 1.
- 7. A host cell transformed with the DNA molecule of claim 2.
- 8. A host cell transformed with the DNA molecule of claim 4.
- 9. A host cell transformed with the DNA molecule of claim 5.
- 10. An isolated DNA molecule having a sequence encoding a truncated BMP receptor protein which is characterized by the ability to bind to BMP in a binding assay, said DNA molecule comprising a DNA sequence selected from the group consisting of:
 - (a) nucleotide 61 to 507 of SEQ ID NO:1;

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- (b) nucleotides encoding amino acids 1 to 149 of SEQ ID NO:2;
- (c) nucleotide 247 to 618 of SEQ ID NO:3;
- (d) nucleotides encoding amino acids 8 to 124 of SEQ ID NO:4; and
- (e) naturally occurring allelic sequences and equivalent degenerative codon sequences of (a), (b), (c) or (d).
 - 11. A host cell transformed with the DNA molecule of claim 10.
- 12. A vector comprising a DNA molecule of claim 10 in operative association with an expression control sequence therefor.
- 13. A method for producing a purified truncated BMP receptor protein, said method comprising the steps of:
- (a) culturing in a culture medium a host cell transformed with a DNA sequence according to claim 10, comprising a nucleotide sequence encoding a truncated BMP receptor protein; and
- (b) recovering and purifying said truncated BMP receptor protein from the culture medium.
- 14. A purified truncated BMP receptor protein comprising the amino acid sequence from amino acid 24 to amino acid 149 of SEQ ID NO: 2.
- 15. A purified truncated BMP receptor protein comprising the amino acid sequence from amino acid 8 to amino acid 124 of SEQ ID NO: 4.
 - 16. A purified truncated BMP receptor protein produced by the steps of:

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(a) culturing in a culture medium a cell transformed with a DNA comprising the nucleotide sequence from nucleotide 61 to 507 of SEQ ID NO: 1; and

- (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid 24 to amino acid 149 of a sequence selected from the group consisting of SEQ ID NO: 2.
 - 17. A purified truncated BMP receptor protein produced by the steps of:
- (a) culturing in a culture medium a cell transformed with a DNA comprising the nucleotide sequence from nucleotide 247 to 618 of SEQ ID NO: 3; and
- (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid 8 to amino acid 124 of a sequence selected from the group consisting of SEQ ID NO: 4.
- 18. A method for producing a truncated BMP receptor protein, said method comprising the steps of:
- (a) culturing in a culture medium a host cell transformed with a DNA sequence encoding a truncated BMP receptor protein, comprising a truncated nucleotide sequence encoding the ligand binding domain of a BMP receptor protein; and
- (b) recovering and purifying said BMP receptor protein from the culture medium.
- 19. A method for producing a truncated BMP receptor protein, said method comprising the steps of:
- (a) culturing in a culture medium a host cell according to claim 11, comprising a truncated nucleotide sequence encoding the ligand binding domain of a BMP receptor protein; and
- (b) recovering and purifying said BMP receptor protein from the culture medium.
- 20. An isolated DNA molecule having a sequence encoding a truncated serine/threonine kinase receptor protein, said DNA molecule comprising a DNA sequence selected from the group consisting of:
 - (a) nucleotide 474 to 836 of SEQ ID NO:5;

(b) nucleotides encoding amino acids 1 to 121 of SEQ ID NO:6;

- (c) nucleotide 80 to 445 of SEQ ID NO:7;
- (d) nucleotides encoding amino acids 1 to 122 of SEQ ID NO:8;
- (e) nucleotide 83 to 445 of SEQ ID NO:9;

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- (f) nucleotides encoding amino acids 1 to 121 of SEQ ID NO:10; and
- (e) naturally occurring allelic sequences and equivalent degenerative codon sequences of (a) through (f).
 - 21. A host cell transformed with the DNA molecule of claim 20.
- 22. A vector comprising a DNA molecule of claim 20 in operative association with an expression control sequence therefor.
- 23. A method for producing a purified truncated serine/threonine kinase receptor protein, said method comprising the steps of:
- (a) culturing in a culture medium a host cell transformed with a DNA sequence according to claim 20, comprising a nucleotide sequence encoding a truncated serine/threonine kinase receptor protein; and
- (b) recovering and purifying said truncated serine/threonine kinase receptor protein from the culture medium.
- 24. A purified truncated serine/threonine kinase receptor protein comprising the amino acid sequence from amino acid 1 to amino acid 121 of SEQ ID NO: 6.
- 25. A purified truncated serine/threonine kinase receptor protein comprising the amino acid sequence from amino acid 1 to amino acid 122 of SEQ ID NO: 8.
- 26. A purified truncated serine/threonine kinase receptor protein comprising the amino acid sequence from amino acid 1 to amino acid 121 of SEQ ID NO: 10.
- 27. A method for producing a truncated serine/threonine kinase receptor protein, said method comprising the steps of:
- (a) culturing in a culture medium a host cell transformed with a DNA sequence encoding a truncated serine/threonine kinase receptor protein, comprising a truncated nucleotide sequence encoding the ligand binding domain of a serine/threonine kinase receptor protein; and

(b) recovering and purifying said truncated serine/threonine kinase receptor protein from the culture medium.

28. A method for producing a truncated serine/threonine kinase receptor protein, said method comprising the steps of:

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- (a) culturing in a culture medium a host cell according to claim 21, comprising a truncated nucleotide sequence encoding the ligand binding domain of a serine/threonine kinase receptor protein; and
- (b) recovering and purifying said truncated serine/threonine kinase receptor protein from the culture medium.

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- 29. A DNA molecule encoding a BMP receptor protein isolated through a method comprising:
- (a) preparing a DNA fragment encoding a ligand binding domain of a known BMP receptor protein as a probe;
- (b) screening a genomic or cDNA library, using the DNA fragment of (a) as a probe;
- (c) isolating a DNA molecule which hybridized to the probe identified in step (b); and
- (d) cloning a DNA molecule encoding a BMP receptor protein from the DNA molecule of step (c).

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- 30. The BMP receptor protein of claim 29, wherein the known BMP receptor protein is CFK1-23a or CFK1-43a.
- 31. An isolated DNA molecule comprising the DNA sequence of CFK1-23a.
- 32. An isolated DNA molecule comprising the DNA sequence of CFK1-43a.
- 33. An isolated DNA molecule comprising the DNA sequence of CFK1-10a.
 - 34. An isolated DNA molecule comprising the DNA sequence of W-101.
 - 35. An isolated DNA molecule comprising the DNA sequence of W-120.

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36. A composition comprising cells transformed with a DNA molecule comprising a DNA sequence encoding one or more BMP receptor proteins.

37. A ligand which binds to a truncated BMP receptor protein, said ligand being selected from the group consisting of an BMP-like monoclonal antibody, a small peptide BMP analogue, and a small organic molecule BMP analogue.

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- 38. The ligand of claim 37, comprising a BMP-like monoclonal antibody.
- 39. The ligand of claim 37, comprising a small peptide BMP analogue.
- 40. The ligand of claim 37, comprising a small organic molecule BMP analogue.
- 41. A pharmaceutical composition comprising a compound first identified for such use as a ligand for the truncated BMP receptor of claim 20.
- 42. The composition of claim 41, comprising an BMP-like monoclonal antibody.
- 43. The composition of claim 41, comprising a small peptide BMP analogue.
- 44. The composition of claim 41, comprising a small organic molecule BMP analogue.
 - 45. A ligand for the truncated BMP receptor protein produced by the method of claim 13, said ligand being selected from the group consisting of an BMP-like monoclonal antibody, a small peptide BMP analogue, and a small organic molecule BMP analogue.
 - 46. The ligand of claim 45, comprising an BMP-like monoclonal antibody.
 - 47. The ligand of claim 45, comprising a small peptide BMP analogue.
 - 48. The ligand of claim 45, comprising a small organic molecule BMP analogue.
 - 49. A pharmaceutical composition comprising a compound first identified for such use as a ligand for the truncated BMP receptor produced by the method of claim 14.
- 50. The composition of claim 49, comprising an BMP-like monoclonal antibody.
 - 51. The composition of claim 49, comprising a small peptide BMP analogue.

52. The composition of claim 49, comprising a small organic molecule BMP analogue.

DESCRIPTION OF THE STREET A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/12 C07K1A CO7K14/71 CO7K14/51 C07K16/28 A61K38/18 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12N C07K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages WO, A, 94 11502 (LUDWIG INSTITUTE FOR CANCER 1-36 P,X RESEARCH, GB) 26 May 1994 see the whole document 3,5,9, P,X WO,A,93 19177 (THE GENERAL HOSPITAL CORP; 20-28 US) 30 September 1993 see the whole document 3,5,9, X DEVELOPMENTAL DYNAMICS, 20-29 vol.196, no.2, February 1993, US pages 133 - 142 HE, W.H. ET AL.; 'Developmental expression of four novel serine/threonine kinase receptors homologous to the activin/transforming growth factor-beta type II receptor family' see the whole document -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. X Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention citation or other special reason (as specified) cannot be considered to involve an inventive step when the document is combined with one or more other such docudocument referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other means document published prior to the international filing date but later than the priority date claimed in the art. "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 3 0. 12. 94 20 December 1994

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X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA., vol.89, December 1992, WASHINGTON US pages 11740 - 11744 CUNNINGHAM, N.S. ET AL.; 'Osteogenin and recombinant bone morphogenetic protein 2B are chemotactic for human monocytes and stimulate transforming growth factor betal m RNA expression' see the whole document	37-52				

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